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(54) Title: PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

(57) Abstract

By this invention, compositions and methods of use related to β -ketoacyl-ACP synthase of special interest are synthases obtainable from *Cuphea* species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

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INTRODUCTION

Field of Invention

The present invention is directed to genes encoding

plant fatty acid synthase enzymes relevant to fatty acid

synthesis in plants, and to methods of using such genes in

combination with genes encoding plant medium-chain

preferring thioesterase proteins. Such uses provide a

method to increase the levels of medium-chain fatty acids

that may be produced in seed oils of transgenic plants.

Background

Higher plants synthesize fatty acids via a common metabolic pathway. In developing seeds, where fatty acids attached to triglycerides are stored as a source of energy for further germination, the fatty acid synthesis pathway is located in the plastids. The first step is the formation of acetyl-ACP (acyl carrier protein) from acetyl-CoA and ACP catalyzed by a short chain preferring condensing enzyme, \$\mathcal{B}\$-ketoacyl-ACP synthase (KAS) III. Elongation of acetyl-ACP to 16- and 18- carbon fatty acids involves the cyclical action of the following sequence of reactions: condensation with a two-carbon unit from malonyl-ACP to form a longer \$\mathcal{B}\$-ketoacyl-ACP (\$\mathcal{B}\$-ketoacyl-ACP synthase), reduction of the

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keto-function to an alcohol (ß-ketoacyl-ACP reductase),
dehydration to form an enoyl-ACP (ß-hydroxyacyl-ACP
dehydrase), and finally reduction of the enoyl-ACP to form
the elongated saturated acyl-ACP (enoyl-ACP reductase). ßketoacyl-ACP synthase I (KAS I), is primarily responsible
for elongation up to palmitoyl-ACP (C16:0), whereas ßketoacyl-ACP synthase II (KAS II) is predominantly
responsible for the final elongation to stearoyl-ACP
(C18:0).

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Genes encoding peptide components of ß-ketoacyl-ACP synthases I and II have been cloned from a number of higher plant species, including castor (Ricinus communis) and Brassica species (USPN 5,510,255). KAS I activity was associated with a single synthase protein factor having an approximate molecular weight of 50 kD (synthase factor B) and KAS II activity was associated with a combination of two synthase protein factors, the 50 kD synthase factor B and a 46 kd protein designated synthase factor A. Cloning and sequence of a plant gene encoding a KAS III protein has been reported by Tai and Jaworski (Plant Physiol. (1993) 103:1361-1367).

The end products of plant fatty acid synthetase activities are usually 16- and 18-carbon fatty acids. There are, however, several plant families that store large

25 amounts of 8- to 14-carbon (medium-chain) fatty acids in their oilseeds. Recent studies with Umbellularia californica (California bay), a plant that produces seed oil rich in lauric acid (12:0), have demonstrated the existence of a medium-chain-specific isozyme of acyl-ACP thioesterase

in the seed plastids. Subsequent purification of the 12:0-ACP thioesterase from Umbellularia californica led to the cloning of a thioesterase cDNA which was expressed in seeds of Arabidopsis and Brassica resulting in a substantial accumulation of lauric acid in the triglyceride pools of these transgenic seeds (USPN 5,512,482). These results and subsequent studies with medium-chain thioesterases from other plant species have confirmed the chain-length-determining role of acyl-ACP thioesterases during de novo fatty acid biosynthesis (T. Voelker (1996) Genetic Engineering, Ed. J. K. Setlow, Vol. 18, pgs. 111-133).

DESCRIPTION OF THE FIGURES

Figure 1. DNA and translated amino acid sequence of Cuphea

hookeriana KAS factor B clone chKAS B-2 are provided.

Figure 2. DNA and translated amino acid sequence of Cuphea
hookeriana KAS factor B clone chKAS B-31-7 are provided.

Figure 3. DNA and translated amino acid sequence of Cuphea
hookeriana KAS factor A clone chKAS A-2-7 are provided.

Figure 4. DNA and translated amino acid sequence of Cuphea
hookeriana KAS factor A clone chKAS A-1-6 are provided.

Figure 5. DNA and translated amino acid sequence of Cuphea
pullcherrima KAS factor B clone cpuKAS B/7-8 are provided.

Figure 6. DNA and translated amino acid sequence of Cuphea
pullcherrima KAS factor B clone cpuKAS B/8-7A are provided.

Figure 7. DNA and translated amino acid sequence of Cuphea
pullcherrima KAS factor A clone cpuKAS A/p7-6A are provided.

Figure 8. Preliminary DNA sequence of Cuphea pullcherrima

KAS factor A clone cpuKAS A/p8-9A is provided.

- Figure 9. DNA and translated amino acid sequence of *Cuphea hookeriana* KASIII clone chKASIII-27 are provided.
- Figure 10. The activity profile for purified cpuKAS B/8-7A using various acyl-ACP substrates is provided.
- 5 Figure 11. The activity profile for purified chKAS A-2-7 and chKAS A-1-6 using various acyl-ACP substrates is provided.
 - Figure 12. The activity profile for purified castor KAS factor A using various acyl-ACP substrates is provided.
- 10 Figure 13. The activity profile for purified castor KAS factor B using various acyl-ACP substrates is provided.

 Figure 14. A graph showing the number of plants arranged according to C8:0 content for transgenic plants containing CpFatB1 versus transgenic plants containing CpFatB1 + chKAS

 15 A-2-7 is provided.
 - Figure 15. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
 - 20 Figure 16. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
 - Figure 17. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.
 - Figure 18. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in

plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.

Figure 19. Graphs showing the %C12:0 in transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 20. Graph showing the relative proportions of C12:0 and C14:0 fatty acids in the seeds of transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 21. Graphs showing the %C18:0 in transgenic plants containing Garm FatB1 (5266) and in seeds of plants resulting from crosses with wild type (X WT) and with lines expressing

15 Ch KAS A-2-7.

Figure 22. The activity profile of Ch KAS A in protein extracts from transgenic plants containing Ch KAS A-2-7.

Extracts were preptreated with the indicated concentrations of cerulenin.

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SUMMARY OF THE INVENTION

By this invention, compositions and methods of use related to ß-ketoacyl-ACP synthase (KAS) are provided. Also of interest are methods and compositions of amino acid and nucleic acid sequences related to biologically active plant synthase(s).

In particular, genes encoding KAS protein factors A and B from *Cuphea* species are provided. The KAS genes are of interest for use in a variety of applications, and may be

used to provide synthase I and/or synthase II activities in transformed host cells, including bacterial cells, such as E. coli, and plant cells. Synthase activities are distinguished by the preferential activity towards longer and shorter acyl-ACPs as well as by the sensitivity towards the KAS specific inhibitor, cerulenin. Synthase protein preparations having preferential activity towards medium chain length acyl-ACPs are synthase I-type or KAS I. KAS I class is sensitive to inhibition by cerulenin at concentrations as low as 1 µM. Synthases having preferential activity towards longer chain length acyl-ACPs are synthase II-type or KAS II. The KAS enzymes of the II-type are also sensitive to cerulenin, but at higher concentrations (50µM). Synthase III-type enzymes have preferential activity towards short chain length acyl-ACPs and are insensitive to cerulenin inhibition.

Nucleic acid sequences encoding a synthase protein may be employed in nucleic acid constructs to modulate the amount of synthase activity present in the host cell, especially the relative amounts of synthase I-type, synthase II-type and synthase III-type activity when the host cell is a plant host cell. In addition, nucleic acid constructs may be designed to decrease expression of endogenous synthase in a plant cell as well. One example is the use of an antisense synthase sequence under the control of a promoter capable of expression in at least those plant cells which normally produce the enzyme.

Of particular interest in the present invention is the coordinate expression of a synthase protein with the

expression of thioesterase proteins. For example, coordinated expression of synthase factor A and a medium-chain thioesterase provides a method for increasing the level of medium-chain fatty acids which may be harvested from transgenic plant seeds. Furthermore, coordinated expression of a synthase factor A gene with plant medium-chain thioesterase proteins also provides a method by which the ratios of various medium-chain fatty acids produced in a transgenic plant may be modified. For example, by expression of a synthase factor A, it, is possible to increase the ratio of C10/C8 fatty acids which are produced in plant seed oils as the result of expression of a thioesterase having activity on C8 and C10 fatty acids.

DETAILED DESCRIPTION OF THE INVENTION

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A plant synthase factor protein of this invention includes a sequence of amino acids or polypeptide which is required for catalyzation of a condensation reaction between an acyl-ACP having a chain length of C2 to C16 and malonyl-ACP in a plant host cell. A particular plant synthase factor protein may be capable of catalyzing a synthase reaction in a plant host cell (for example as a monomer or homodimer) or may be one component of a multiple peptide enzyme which is capable of catalyzing a synthase reaction in a plant host cell, i.e. one peptide of a heterodimer.

Synthase I (KAS I) demonstrates preferential activity towards acyl-ACPs having shorter carbon chains, C2-C14 and is sensitive to inhibition by cerulenin at concentrations of 1µM. Synthase II (KAS II) demonstrates preferential

activity towards acyl-ACPs having longer carbon chains, C₁₄-C₁₆, and is inhibited by concentrations of cerulenin (50µM). Synthase III demonstrates preferential activity towards acyl-CoAs having very short carbon chains, C₂ to C₆, and is insensitive to inhibition by cerulenin.

Synthase factors A and B, and synthase III proteins obtained from medium-chain fatty acid producing plant species of the genus Cuphea are described herein. As described in the following Examples, synthase A from C. hookeriana is naturally expressed at a high level and only in the seeds. C. hookeriana synthase B is expressed at low levels in all tissues examined. Expression of synthase A and synthase B factors in E. coli and purification of the resulting proteins is employed to determine activity of the various synthase factors. Results of these analyses indicate that synthase factor A from Cuphea hookeriana has the greatest activity on 6:0-ACP substrates, whereas synthase factor B from Cuphea pullcherrima has greatest activity on 14:0-ACP. Similar studies with synthase factors A and B from castor demonstrate similar activity profiles between the factor B synthase proteins from Cuphea and The synthase A clone from castor, however, demonstrates a preference for 14:0-ACP substrate.

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Expression of a Cuphea hookeriana KAS A protein in

transgenic plant seeds which normally do not produce mediumchain fatty acids does not result in any detectable
modification of the fatty acid types and contents produced
in such seeds. However, when Cuphea hookeriana KAS A
protein is expressed in conjunction with expression of a

medium-chain acyl-ACP thioesterase capable of providing for production of C8 and C10 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain

5 thioesterase alone are observed. In addition, where significant amounts of C8 and C10 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C8 and C10 fatty acids

10 that are obtained. For example, an increased proportion of C10 fatty acids may be obtained by co-expression of Cuphea hookeriana ChFatB2 thioesterase and a chKAS A synthase factor proteins.

Furthermore, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a medium-chain acyl-ACP thioesterase capable of providing for production of C12 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are also observed. In addition, where significant amounts of C12 and C14 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C12 and C14 fatty acids that are obtained. For example, an increased proportion of C12 fatty acids may be obtained by co-expression of UC FatB1 thioesterase and a chKAS A synthase factor proteins.

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However, when Cuphea hookeriana KAS A protein is expressed in conjunction with the expression of a long-chain

acyl-ACP thioesterase capable of providing for production of C18 and C18:1 fatty acids in plant seed oils, no effect on the production of long chain fatty acids was observed.

Furthermore, when plants transformed to express a long chain acyl-ACP thioesterase from mangosteen (GarmFatAl, U.S. Patent Application No. 08/440,845), which preferentially hydrolyzes C18:0 and C18:1 fatty acyl-ACPs, are crossed with nontransformed control plants, a significant reduction in the levels of C18:0 is obtained. Similar reductions are also observed in the levels of C18:0 in the seeds of plants resulting from crosses between plants transformed to express the GarmFatAl and plants expressing the Cuphea hookeriana KAS A protein.

Thus, the instant invention provides methods of increasing and/or altering the medium-chain fatty acid 15 compositions in transgenic plant seed oils by co-expression of medium-chain acyl-ACP thioesterases with synthase factor proteins. Furthermore, various combinations of synthase factors and medium-chain thioesterases may be achieved depending upon the particular fatty acids desired. For 20 example, for increased production of C14 fatty acids, synthase protein factors may be expressed in combination with a C14 thioesterase, for example from Cuphea palustris or nutmeg may be employed (WO 96/23892). In addition, thioesterase expression may be combined with a number of different synthase factor proteins for additional effects on medium-chain fatty acid composition.

Synthases of use in the present invention include modified amino acid sequences, such as sequences which have

been mutated, truncated, increased and the like, as well as such sequences which are partially or wholly artificially synthesized. The synthase protein encoding sequences provided herein may be employed in probes for further screening or used in genetic engineering constructs for transcription or transcription and translation in host cells, especially plant host cells. One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover synthases and/or synthase nucleic acid sequences from other sources. Typically, a homologously related nucleic acid sequence will show at least about 60% homology, and more preferably at least about 70% homology, between the R. communis synthase and the given plant synthase of interest, excluding any deletions which may be present. Homology is determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions.

Recombinant constructs containing a nucleic acid

sequence encoding a synthase protein factor or nucleic acid

sequences encoding a synthase protein factor and a mediumchain acyl-ACP thioesterase may be prepared by methods well

known in the art. Constructs may be designed to produce

synthase in either prokaryotic or eukaryotic cells. The

increased expression of a synthase in a plant cell,

particularly in conjunction with expression of medium-chain
thioesterases, or decreasing the amount of endogenous

synthase observed in plant cells are of special interest.

Synthase protein factors may be used, alone or in combination, to catalyze the elongating condensation reactions of fatty acid synthesis depending upon the desired result. For example, rate influencing synthase activity may reside in synthase I-type, synthase II-type, synthase III-type or in a combination of these enzymes. Furthermore, synthase activities may rely on a combination of the various synthase factors described herein.

Constructs which contain elements to provide the

transcription and translation of a nucleic acid sequence of interest in a host cell are "expression cassettes".

Depending upon the host, the regulatory regions will vary, including regions from structural genes from viruses, plasmid or chromosomal genes, or the like. For expression in prokaryotic or eukaryotic microorganisms, particularly unicellular hosts, a wide variety of constitutive or regulatable promoters may be employed. Among transcriptional initiation regions which have been described are regions from bacterial and yeast hosts, such as E. coli,

B. subtilis, Saccharomyces cerevisiae, including genes such as &-galactosidase, T7 polymerase, trp-lac (tac), trp E and the like.

An expression cassette for expression of synthase in a plant cell will include, in the 5' to 3' direction of

25 transcription, a transcription and translation initiation control regulatory region (also known as a "promoter") functional in a plant cell, a nucleic acid sequence encoding a synthase, and a transcription termination region.

Numerous transcription initiation regions are available

which provide for a wide variety of constitutive or regulatable, e.g., inducible, transcription of the desaturase structural gene. Among transcriptional initiation regions used for plants are such regions associated with cauliflower mosaic viruses (35S, 19S), and structural genes such as for nopaline synthase or mannopine synthase or napin and ACP promoters, etc. The transcription/ translation initiation regions corresponding to such structural genes are found immediately 5' upstream to the respective start codons. Thus, depending upon the intended use, different promoters may be desired.

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Of special interest in this invention are the use of promoters which are capable of preferentially expressing the synthase in seed tissue, in particular, at early stages of seed oil formation. Examples of such seed-specific promoters include the region immediately 5' upstream of a napin or seed ACP genes such as described in USPN 5,420,034, desaturase genes such as described in Thompson et al (Proc. Nat. Acad. Sci. (1991) 88:2578-2582), or a Bce-4 gene such as described in USPN 5,530,194. Alternatively, the use of the 5' regulatory region associated with the plant synthase structural gene, i.e., the region immediately 5' upstream to a plant synthase structural gene and/or the transcription termination regions found immediately 3' downstream to the plant synthase structural gene, may often be desired. general, promoters will be selected based upon their expression profile which may change given the particular application.

In addition, one may choose to provide for the transcription or transcription and translation of one or more other sequences of interest in concert with the expression or anti-sense of the synthase sequence, particularly medium-chain plant thioesterases such as described in USPN 5,512,482, to affect alterations in the amounts and/or composition of plant oils.

When one wishes to provide a plant transformed for the combined effect of more than one nucleic acid sequence of interest, a separate nucleic acid construct may be provided for each or the constructs may both be present on the same plant transformation construct. The constructs may be introduced into the host cells by the same or different methods, including the introduction of such a trait by crossing transgenic plants via traditional plant breeding methods, so long as the resulting product is a plant having both characteristics integrated into its genome.

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Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformed cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species into which the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

The manner in which the DNA construct is introduced into the plant host is not critical to this invention. Any method which provides for efficient transformation may be employed. Various methods for plant cell transformation include the use of Ti- or Ri-plasmids, microinjection, electroporation, liposome fusion, DNA bombardment or the like. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses A. tumefaciens or A. rhizogenes as a mode for transformation, although the T-DNA borders may find use with other modes of transformation.

The expression constructs may be employed with a wide variety of plant life, particularly plant life involved in the production of vegetable oils. These plants include, but are not limited to rapeseed, peanut, sunflower, safflower, cotton, soybean, corn and oilseed palm.

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For transformation of plant cells using Agrobacterium, explants may be combined and incubated with the transformed Agrobacterium for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

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Example 1 Cuphea KAS Factor A and B Gene Cloning

Total RNA isolated from developing seeds of Cuphea 10 hookeriana and Cuphea pullcherrima was used for cDNA synthesis in commercial 1-based cloning vectors. For cloning each type of KAS gene, approximately 400,000-500,000 unamplified recombinant phage were plated and the plaques transferred to nitrocellulose. For KAS factor B cloning from C. hookeriana, a mixed probe containing Brassica napus 15 KAS factor B and Ricinus communis (Castor) KAS factor B radiolabeled cDNA's was used. Similarly, a mixed probe containing Brassica napus KAS factor A and Ricinus communis KAS factor A cDNA clones was used to obtain C. hookeriana 20 KAS factor A genes. For KASIII, a spinach KASIII cDNA clone obtained from Dr. Jan Jaworski was radiolabeled and used as a probe to isolate a KASIII clone from C. hookeriana. For KAS B and KAS A cloning from C. pullcherrima, C. hookeriana KAS B and KAS A genes chKAS B-2 and chKAS A-2-7 (see below) were radiolabeled and used as probes.

DNA sequence and translated amino acid sequence for Cuphea KAS clones are provided in Figures 1-9. Cuphea hookeriana KAS factor B clones chKAS B-2 and chKAS B-31-7 are provided in Figures 1 and 2. Neither of the clones is full length. Cuphea hookeriana KAS Factor A clones chKAS A-2-7 and chKAS A-1-6 are provided in Figures 3 and 4. chKAS A-2-7 contains the entire encoding sequence for the KAS factor protein. Based on comparison with other plant synthase proteins, the transit peptide is believed to be represented in the amino acids encoded by nucleotides 125-466. chKAS A-1-6 is not a full length clone although some transit peptide encoding sequence is present. Nucleotides 1-180 represent transit peptide encoding sequence, and the mature protein encoding sequence is believed to begin at nucleotide 181.

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Cuphea pullcherrima KAS factor B clones cpuKAS B/7-8 and cpuKAS B/8-7A are provided in Figures 5 and 6. Both of the clones contain the entire encoding sequences for the KAS 15 factor B proteins. The first 35 amino acids of cpuKAS B/7-8 are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide The first 39 amino acids of cpuKAS B/8-7A are believed to represent the transit peptide, with the mature protein 20 encoding sequence beginning at nucleotide 209. Cuphea pullcherrima KAS factor A clones cpuKAS A/p7-6A and cpuKAS A-p8-9A are provided in Figures 7 and 8. Both of the clones contain the entire encoding sequences for the KAS factor A proteins. Translated amino acid sequence of cpuKAS A/p7-6A is provided. The mature protein is believed to begin at the lysine residue encoded 595-597, and the first 126 amino acids are believed to represent the transit peptide. The DNA sequence of KAS A clone cpuKAS A-p8-9A is preliminary.

Further analysis will be conducted to determine final DNA sequence and reveal the amino acid sequence encoded by this gene.

DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 is provided in Figure 9. The encoding sequence from nucleotides 37-144 of chKASIII-27 are believed to encode a transit peptide, and the presumed mature protein encoding sequence is from nucleotides 145-1233.

Deduced amino acid sequence of the C. hookeriana KAS 10 factor B and KAS factor A cDNA's reveals strong homology to the Brassica napus and Ricinus communis clones previously reported. The C. hookeriana KAS factor B clone is more homologous to the Ricinus and Brassica KAS factor B clones (94% and 91% respectively) than it is to the Ricinus and Brassica KAS factor A clones (60% for both). Furthermore, the C. hookeriana KAS factor A clone is more homologous to the Ricinus and Brassica KAS factor A clones (85% and 82% respectively) than it is the Ricinus and Brassica KAS factor 20 B clone (60% for both). The C. hookeriana KAS factor B cDNAs designated as chKAS B-2 and chKAS B-31-7 are 96% identical within the mature portion of the polypeptide. Similarly, the deduced amino acid sequence of the mature protein regions of the C. hookeriana KAS factor A clones chKAS A-2-7 and chKAS A-1-6 are 96% identical. The C. pullcherrima KAS clones also demonstrate homology to the R. communis and Brassica napus KAS clones. The mature protein portion of all of the KAS factor A family members in the different Cuphea species are 95% identical. Similarly the

mature protein portion of the KAS factor B genes in Cuphea are also 95-97% identical with each other. However there is only approximately 60% sequence identity between KAS factor B and KAS factor A clones either within the same or different species of Cuphea.

Example 2 Levels and Patterns of Expression

To examine tissue specificity of KAS expression in Cuphea hookeriana, Northern blot analysis was conducted using total RNA isolated from seed, root, leaf and flower tissue. Two separate but identical blots were hybridized with either chKAS B-31-7 or chKAS A-2-7 coding region probes. The data from this RNA blot analysis indicate that KAS B is expressed at a similar level in all tissues examined, whereas KAS A expression is detected only in the These results also demonstrate a different level of expression for each of the synthases. KAS A is an abundant message, whereas KAS B is expressed at low levels. Furthermore, even under highly stringent hybridization conditions (65_C, 0.1 X SSC, 0.5% SDS), the KAS A probe hybridizes equally well with two seed transcripts of 2.3 and The larger hybridizing band is likely the transcript of the KAS A-2-7 gene since the size of its cDNA is 2046bp, and the number of clones obtained from cDNA screening corresponds well with the apparent mobility of the mRNA and its abundance on the blot.

Example 3 Expression of Plant KAS Genes in E.coli

DNA fragments encoding the mature polypeptide of the Cuphea hookeriana KAS A cDNAs and the Cuphea pullcherrima

5 KAS B cDNAs were obtained by PCR and cloned into a
QIAexpress expression vector (Qiagene). Experimental
conditions for maximum level of expression were determined
for all of these clones and the parameters for highest level
of soluble fraction were identified. Cells are grown in

10 ECLB media containing 1M sorbitol and 2.5 mM betaine
overnight and subcultured as a 1:4 dilution in the same
medium. Cells are then grown for 2 hours (to approximately
.6-.8 O.D.) and induced with 0.4 mM IPTG and allowed to grow
for 5 more hours.

Enzyme activity of the affinity purified recombinant enzymes obtained from over-expression of the chKAS A-2-7 and cpuKAS B/8-7A clones was measured using a wide range of acyl-ACP substrates (6:0- to 16:1-ACP). The activity profile for cpuKAS B/8-7A is provided in Fig.10. The data demonstrate that the enzyme is active with all acyl-ACP substrates examined, although activity on 6:0 to 14:0-ACP substrates is substantially greater than the activity on 16:0 and 16:1 substrates.

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The activity profile of the *C. hookeriana* KAS A clones

25 chKAS A-2-7 and chKAS A-1-6 is provided in Figure 11. The *C. hookeriana* KAS A clones are most active with C:6, and have the least activity with C:16:0 substrates. However, the activity of this clone on even the preferred C6:0 substrate

is 50 fold lower than the activity of the *C. pullcherrima* KAS B clones.

A fragment containing the mature protein encoding portion of a R. communis KAS factor A clone was also cloned into a QIAexpress expression vector, expressed in E. coli and the enzyme affinity purified as described above. The activity profile for castor KAS A is provided in Figure 12. Highest activity is observed with C14:0 substrates, although some activity is also seen with C6:0 and C16:1. 10 comparison, the activity profile obtained from purified R. communis KAS factor B also using the QIAexpress expression system is provided in Figure 13. The KAS B clone demonstrates substantially higher levels of activity (10 fold and higher) than the R. communis KAS A clone. preference of the KAS factor B for 6:0- to 14:0-ACP substrates is consistent with the previous observations that this protein provides KAS I activity.

Example 4 KAS and TE Expression in Transgenic Seed

Dehesh et al. (1996) Plant Physiol. 110:203-210) and the chKAS A-2-7 were PCR amplified, sequenced, and cloned into a napin expression cassette. The napin/cp FatBl and the napin/KAS A-2-7 fusions were ligated separately into the binary vector pCGN1558 (McBride and Summerfelt (Pl.Mol.Biol. (1990) 14:269-276) and transformed into A. tumefaciens, EHA101. The resulting CpFatBl binary construct is pCGN5400 and the chKAS A-2-7 construct is pCGN5401. Agrobacterium mediated transformation of a Brassica napus canola variety

was carried out as described by Radke et al. (Theor. Appl. Genet. (1988) 75:685-694; Plant Cell Reports (1992) 11:499-505). Several transgenic events were produced for each of the pCGN5400 and pCGN5401 constructs.

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A double gene construct containing a napin/cpFatB1 expression construct in combination with a napin/chKAS A-2-7 expression construct was also assembled, ligated into a binary vector and used for co-cultivation of a canola Brassica variety. The binary construct containing the chFatB1 and chKAS A-2-7 expression constructs is pCGN5413.

Fatty acid analysis of 26 transgenic lines containing chKAS A-2-7 (5401 lines) showed no significant changes in the oil content or profile as compared to similar analyses of wild type canola seeds of the transformed variety.

15 Fatty acid analysis of 36 transgenic lines containing cpFatB1 (5400 lines) showed increased levels of C:8 and C:10 in transgenic seeds. The highest level of C:8 observed in a pool seed sample was 4.2 mol%. The C:10 levels were between 30 and 35% of the C:8 content. Fatty acid analysis of 25 - 20 transgenic lines containing the TE/KAS A tandem (5413 lines) demonstrated an overall increase in both C:8 and C:10 levels relative to those observed with TE containing lines (5400) In lines containing the cpFatB1 construct alone, the average level of C:8 average were 1.5 mol%, whereas the C:8 average levels in TE/KAS A tandem containing lines was 2.37 The ratio of C:8 to C:10 remained constant in both populations. The number of transgenic events relative to the C:8 content are presented in Figure 14. These data show that the transgenic events with tandem TE/KAS A construct

yield more lines with higher levels of C:8 than those events with single TE construct. For example, several lines containing nearly 7 mole% C8 were obtained with the TE/KAS A pCGN5413 construct, whereas the highest C8 containing line from the pCGN5400 TE alone transformation contained 4.2 mole% C8.

Half seed analysis of the T3 generation of transgenic canola plants expressing a ChFatB2 (*C. hookeriana* thioesterase; Dehesh et al. (1996) The Plant Journal 9:167-172) indicate that these plant can accumulate up to 22 weight% (33 mol%) of 8:0 and 10:0 fatty acids (4804-22-357). Segregation analysis shows that these transformants contain two loci and that they are now homozygous. Selected plants grown from these half seeds were transferred into the greenhouse and later crossed with T1 transformants that had been transformed with either Cuphea hookeriana KAS A (5401) alone or KAS A/CpFatB1 double constructs (5413).

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Fatty acid analysis of several events resulting from the crosses between transgenic lines containing ChFatB2

20 (4804-22-357) and chKAS A-2-7 (5401-9), reveal an increase in the ratio of C:10/C:8 levels (Figure 15). This C:10/C:8 ratio in nearly all of the transgenic events containing ChFatB2 TE alone fluctuates between 3 and 6, whereas in the F1 generation of transgenic containing both the TE and the KAS A-2-7, the ratio can be as high as 22. This increase in C:10 levels is accompanied by an increase in the total C:8 and C:10 content (Figure 16). The sum of the C:8 and C:10 fatty acids in the heterozygous F1 lines is as high as those in the homozygous parent line (4804-22-357), whereas the

heterozygous lines usually contain substantially less C:8 and C:10 than the homozygous lines.

Similar results were observed in F1 generation seeds resulting from crosses performed between 4804-22-357 (ChFatB2) and the 5413-17 event (CpFatB1 and chKAS A-2-7 tandem). Levels of C:8 and C:10 in the 5413-17 line were 6.3 and 2.8 mol% respectively. Data presented in Figure 17 show that there is shift towards C:10 fatty acids as was observed with the 4804-22-357 (ChFatB2) x 5401-9 (chKAS A-2-7) crosses. Furthermore, Figure 18 indicates the presence 10 of two separate populations of heterozygotes. containing approximately 9-11 weight percent C:10 + C:8 are believed to represent offspring containing a single copy of the ChFatB1 TE gene and no copies of the CpFatB1 and chKAS A genes from 5413. Those plants containing approximately 15-15 20 weight percent C:10 + C:8 are believed to represent the heterozygotes containing a single ChFatB1 TE gene as well as the CpFatB1 and chKAS A genes from 5413. Thus, the level of the C:10 + C:8 fatty acids does not decrease to 50% of that 20 detected in parent lines when a copy of the ChKAS A gene is present.

To further characterize the chain length specificity of the Cuphea hookeriana KAS A enzyme, crosses between transgenic Brassica napus lines containing a California Bay (Umbellularia californica) 12:0 specific thioesterase, Uc FatB1 (USPN 5,344,771) and chKAS A-2-7 (5401-9) were made. Half seed analysis of transgenic plants containing Uc fatB1 have previuosly indicated that these plants can accumulate up to 52 mol% C12:0 in the seed oil of homozygous dihaploid

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lines (LA86DH186). Crosses between the line LA86DH186 and untransformed control *Brassica* demonstrated a decrease in the C12:0 levels.

However, crosses between LA86DH186 and the 5401-9

5 hemizygous line led to an accumulation of up to 57 mol%

C12:0 in the seed oil of F1 progeny (Figure 19).

Interestingly, in crosses with LA86DH186 x untransformed control line and LA86DH186 x 5401-9, levels of C14:0 in the seeds of the F1 progeny decreased to 50% of the levels

10 obtained in homozygous LA86DH186 lines (Figure 20).

Furthermore, increases in the proportion of C12:0 fatty acid resulted in a substantial decline in the proportions of all the long-chain fatty acyl groups (C16:0, C18:0, C18:2, and C18:3). These results indicate that the ChKAS A-2-7 is an enzyme with substrate specificity ranging from C6:0 to C10:0-ACP, and that its over-expression ultimately reduces the longer chain acyl-ACP pools.

Further evidence is obtained in support of the chain length specificity of the ChKAS A-2-7 in crosses of the 5401-9 line with a transgenic line (5266) expressing an 18:1/18:0 TE from Garcinia mangostana (GarmFatA1, US patent application No. 08/440,845). Transgenic Brassica line 5266 has been shown to accumulate up to 24 mol% C18:0 in the seed oil of homozygous lines (Figure 21). However, in the seed oil of F1 progeny of crosses between 5266 and 5401-9 levels of C18:0 were reduced to approximately 12 mol%. Furthermore, levels of C16:0 generated from these crosses was similar to the levels obtained from the seed oil of nontransgenic control plants.

Example 5 In vitro Analysis of Plant KAS Enzymes

Seed extracts were prepared from developing seeds of nontransgenic controls or transgenic Brassica expressing chKAS A-2-7 as described in Slabaugh et al. (Plant Journal. 5 1998 in press) and Leonard et al. (Plant Journal, 1998, in press). In vitro fatty acid synthesis assays were performed as described by Post-Beittenmiller (J. Biol. Chem. (1991), 266:1858-1865). Extracts were concentrated by ammonium sulfate precipitation and desalting using P-6 columns (Bio-Rad, Hercules, CA). Reactions (65µ1) contained 0.1M Tris/HCl (pH 8.0), 1 mM dithiothreitol, 25 mM recombinant spinach ACP1, 1 mM NADH, 2 mM NADPH, 50 μM malonyl-CoA, 10 μM [1-14C]acetyl-CoA (50 mCi/mmol), 1mg/ml BSA, and 0.25 mg/ml seed protein. Selected seed extracts were preincubated with cerulenin at 23°C for 10 min. Reaction products were separated on an 18% acrlamide gel containing 2.25M urea, electroblotted onto to nitrocellulose and quntitated by phosporimaging using Image QuaNT software (Molecular Dynamics, Sunnyvale, CA). Authentic acyl-ACPs were run in parallel, immunoblotted and finally detected by anti-ACP serum to confirm fatty acid chain lengths.

The results (Figure 22) indicate that the fatty acid synthesis capabilities of transgenic Brasica (5401-9) seed extracts was greater than that obtained from in the nontransgenic controls as measured by the relative abundance of C8:0- and C10:0-ACP at all time points tested. In addition, pretreatment of the extracts with cerulenin, markedly reduced the synthesis of longer chain fatty acids in both the transgenic and nontransgenic control seed

- 20

extracts. However, the extension of the spinach-ACP was much less inhibited in the seed extracts from the transgenic lines than in the seed extracts of nontransgenic control Brassica.

These data further support that Ch KAS A-2-7 is a condensing enzyme active on medium chain acyl-ACPs, and that expression of this enzyme in plants results in enlarged substrate pools to be hydrolyzed by medium-chain specific thioesterases. Furthermore, these data suggest that chKAS A-2-7 also is a cerulenin-resistant condensing enzyme.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains.

- 15 All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.
- Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

MISSING UPON TIME OF PUBLICATION

- 13. The construct of Claim 5 wherein said encoding sequence is cpuKAS A/p8-9A.
- 14. The construct of Claim 5 wherein said encoding sequence is chKASIII-27.
- 15. An improved method for producing medium-chain fatty acids in transgenic plant seeds by expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant,

the improvement comprising expression of a plant synthase

factor protein heterologous to said transgenic plant in

conjunction with expression of said plant medium-chain

thioesterase, whereby the percentage of medium-chain fatty

acids produced in seeds expressing both a plant synthase factor

protein and a plant medium-chain thioesterase protein is

increased as compared to the percentage of medium-chain fatty

- acids produced in seeds expressing only said plant medium-chain thioesterase protein.
- 16. The method of Claim 15 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 20 17. The method of Claim 15 wherein said medium-chain thioesterase protein is a CpFatBl protein.
 - 18. The method of Claim 15 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 25 19. The method of Claim 15 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 20. The method of Claim 19 wherein said synthase factor A protein is from a Cuphea species.

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- 21. The method of Claim 20 wherein said Cuphea species is C. hookeriana or C. pullcherrima.
- 22. A method of altering the medium-chain fatty acid composition in plant seeds expressing a heterologous plant medium-chain preferring thioesterase, wherein said method comprises

providing for expression of a plant synthase factor protein heterologous to said transgenic plant in conjunction with expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant, whereby the composition of medium-chain fatty acids produced in said seeds is modified as compared to the composition of medium-chain fatty acids produced in seeds expressing said plant medium-chain thioesterase protein in the absence of expression of said plant synthase factor protein.

- 23. The method of Claim 22 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 24. The method of Claim 22 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
- 25. The method of Claim 22 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
 - 26. The method of Claim 22 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 27. The method of Claim 26 wherein said synthase factor A protein is from a Cuphea species.
 - 28. The method of Claim 27 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

- 29. The method of Claim 22 wherein said fatty acid composition is enriched for C10 fatty acids.
- 30. The method of Claim 22 wherein said fatty acid composition is enriched for C12 fatty acids.
- 31. The method of Claim 22 wherein said fatty acid composition is enriched for at least one medium chain fatty acid and at least one other medium chain fatty acid is decreased.
- 32. The method of Claim 31 wherein said enriched fatty 10 acid is C12 and said decreased fatty acid is C14.

48	96	144	192	240	288	336	384
66C 61y	AAG Lys	GGT Gly	CAC His	666 61y	TCA Ser	GCT Ala	ACT Thr
CCG	TCC Ser	GGT Gly	GGT Gly	ATG Met	TAT	GCC	66C 61Y
CCC	CTC	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT	GGA Gly
GAT Asp	CGC	GGA Gly	GAG Glu	ACA	CCA	CAT	GCT Ala
GTG Val	GAC Asp	ACA Thr	ATC Ile	ATT Ile	66C 61y	TTC	ATT Ile
CTA Leu	GCC	GGA Gly	CTT Leu	GCC Ala	ATG Met	TGC Cys	ATG
gaa glu	GGT	GTC Val	TCT Ser	TAT Tyr	CTC Leu	TAC Tyr	CTT
CTA	CTC	CTG	CAG	CCC	GGT Gly	AAC Asn	GAT
GCT Ala	GAT Asp	GTG Val	GTT Val	ATC Ile	TTT Phe	TCC	GCT
GCC	GCC Ala	GGA Gly	$^{\tt GGG}_{\tt G1Y}$	TTC	GAA Glu	ACT Thr	GAG Glu
GCG Ala	CGA Arg	GCC	GAC Asp	TTC Phe	ATC	GCC	GGT Gly
GTG Val	GCA	AGA Arg	TCT Ser	CCT	GCT Ala	TGT	CGT
GCG Ala	TCG Ser	GAG Glu	TTC	ACC	CTC	GCA	CGC
ACC Thr	AAT Asn	AAG Lys	GTC Val	ATC	CTG	ACT Thr	ATC Ile
JCC	AGG Arg	GAC Asp	ACT Thr	AAA Lys	GCC	JCC	CAT
AGC	TGC	ATC Ile	CTG	CGG Arg	TCT	ATT	AAT

FIGURE 1 1 OF 4

432	480	528	576	624	672	720	768
AGG Arg	TGG Trp	TTG	ATT Ile	ACT	AGC Ser	GCT Ala	ATC Ile
TGC Cys	CCC	GTG Val	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala
GCT Ala	AGG Arg	GGA G1Y	CCG	CAC	GAG Glu	ATA Ile	AAT
GTG Val	TCT Ser	GCT Ala	GCA	TAT Tyr	ATT	TAC	ATA
TTT Phe	GCC Ala	GGT Gly	GGA G1y	GCT Ala	TGC	AAT Asn	GAG Glu
GGC G1y	ACT	GAA Glu	CGA Arg	gat Asp	TCT Ser	GTC Val	GCC
634 617	CAG Gln	GGT Gly	aga Arg	TGT Cys	TCT Ser	GAG Glu	CTC Leu RE 1 F4
TTG	CCG	ATG Met	ATG Met	AAC Asn	GTC Val	GAA Glu	GAT CTC Asp Leu FIGURE 1 2 OF 4
666 617	gac Asp	GTG Val	GCA Ala	ATC	GGT Gly	CCT	666 Gly
ATT Ile	gat Asp	TTT Phe	CAT His	GCA Ala	CTT Leu	TCA	GCT Ala
CCA	AAC Asn	GGT Gly	GAA Glu	GGT G1Y	GGT Gly	GTC Val	CTA
ATT Ile	AGG Arg	GAT Asp	TTG Leu	GGA Gly	gat Asp	66C G1y	ACT
ATC Ile	CAA Gln	CGT Arg	AGC Ser	TTG Leu	GCT Ala	GCT Ala	Ser
GCA Ala	TCT	GAC Asp	GAG Glu	TAT Tyr	AGG Arg	gat Asp	ACT
GCC	TTG Leu	aaa Lys	ATG Met	GAG Glu	CCA	GAA Glu	GCG Ala
GAG Glu	GCT Ala	GAT ASD	GTG Val	GCA	gat Asp	CTT Leu	CAT

816	864	912	096	1008	1056	1116
aag Lys	ATA Ile	AAT	AAG Lys	TTT Phe	e E	CCCATITICAC AAGGIACITG ICAITGAGAA TACGGAITAT GGACITGCAG AGIAAITITCC CCATGITIGI CGGAAGAGCA TAITACCACG GITGICGIC AAACCCAITI AGGAIACIGI
ACT	GCT	ATT	AAC	GGA G1Y	CCA TGATTA Pro	GTAA
GCA Ala	GAA Glu	AGC	GCC Ala	TTC Phe		'AG A 'T'T A
AAT	CTT Leu	CCC Pro	GTT Val	TCA Ser	AAG Lys	TTGC
ATT Ile	$_{\rm GIY}^{\rm GGT}$	CAT	ACT	AAT Asn	TTC	GGAC
AAA Lys	GGA Gly	CTT Leu	GAC Asp	TCG Ser	GCT	TTAT
ATC Ile	TCT Ser	TGG Trp	TTC	ATC	TCG Ser	TACGGATTAT GTTGTCCGTC
GAT Asp	GCA Ala	66C 61y	GAG Glu	GCG	TTC	A TAC
AAG Lys	GGA Gly	ACC	GTG Val	GTT Val	GCT Ala	TCATTGAGAA TATTACCACG
ACA Thr	CTT	AAC	TCG	AAC Asn	GTG Val	CATTC
AAC Asn	TGT Cys	ATA Ile	CCA	GTT Val	GTC Val	55 F.C.
AAG Lys	CAC	GGA G1y	GAG Glu	gaa g1u	TCA	PACT'T AGAGO
TTC	GGA	AAG Lys	CCT	CAC His	AAC Asn	AAGG1
GTT Val	ATC Ile	ATT Ile	AAT Asn	CAA Gln	CAC	CAC A
AAG Lys	ATG Met	ACT Thr	TTC	CAG Gln	GGC Gly	CCCATTTCAC AAGGTACTTG CCATGTTTGT CGGAAGAGCA
AAG Lys	TCA	GCG Ala	CAA Gln	AAG Lys	GGA Gly	CCC?

FIGURE 1

1348	AA	ACTTTTGTTT GTATTGGAAA GGAAGTGCCG TCTCAAAAAA AAAAAAAAA AA	TCTCAAAAAA	GGAAGTGCCG	GTATTGGAAA	ACTTTTGTTT
1296	TGAAATTATA TITATITIAT CITAGAAAGG TCAAATAAGA TITITITITA CCICTGTAAA 1296	TTTTGTTTTA	TCAAATAAGA	CTTAGAAAGG	TTTATTTAT	TGAAATTATA
1236	TCTATGTAAT AAAACTAAGG ATTATTAATT TCCCTTTTAA TCCTGTCTCC AGTTTGAGCA 1236	TCCTGTCTCC	TCCCTTTTAA	ATTATTAATT	AAAACTAAGG	TCTATGTAAT

S

ACC GNG GTG Thr Xxx Val> CCG GGC TGC AGG AAT TCG GCA Pro Gly Cys Arg Asn Ser Ala> GGC TCC GAC GTC GAC TCT Gly Ser Asp Val Asp Ser> ATC AGC TTA ATC GAC Ile Ser Leu Ile Asp> 270 GGG AAG AAC GAC AGG AGG CTC Gly Lys Asn Asp Arg Arg Leu> GGC CAG 2 Gly Gln 3 90 330 30 TCC 230 TGG AGC Trp Ser 66C 61y 180 130 999 Gly TCC AAG TTC CCC ACC AGG TTC Ser Lys Phe Pro Thr Arg Phe 80 320 20 AGC Ser TCC GTA TTC Ser Val Phe GGC GAG AGC Gly Glu Ser 220 AAA TTA ACC CTC ACT AAA GGG AAC AAA Lys Leu Thr Leu Thr Lys Gly Asn Lys GCG GCC GCT CTA GAA CTA GTG GAT CCC Ala Ala Ala Leu Glu Leu Val Asp Pro 260 ACG GGA TAC ATC GAC Thr Gly Tyr Ile Asp 170 120 70 310 GGC ATG GGC CTC GTC Gly Met Gly Leu Val TAC GAA AAG CTC CTC TCC TYr Glu Lys Leu Leu Ser Sequence Range: 1 to 1704 210 160 110 250 TTC AAC GCG P Phe Asn Ala 1 TTC GAC GCT Phe Asp Ala 300 200 CGA GCC (Arg Ala (100 TAT CGC GGA Gly

190

140

FIGURE 1/5

GAC GAT TGC CTC CGC TAC TGC ATT GTC GCC GGG AAG AAG GCT CTC GAA Asp Asp Cys Leu Arg Tyr Cys Ile Val Ala Gly Lys Lys Ala Leu Glu>

·	AGA Arg>	0.0	TCT Ser>	480	CCG Pro>	GCC Ala>		TGT Cys>		CGA Arg>	0	ATT Ile>
380	GAG	43	TTC		TCC	CTT Leu		GCA Ala	620	CGC	670	ATC Ile
•••	AAG Lys		GTC Val		ATC Ile	CTG Leu	570	ACT	Ψ	ATC		GCA
	GAT Asp		ACC	470	AAG Lys	520 GCT CT Ala Le		TCA		CAT His		GCT Ala
370	ATT Ile	420	CTA	4	CGG	TCT Ser		ATT	610	AAT Asn	* 099	GAG Glu
'n	AAG Lys		66C 61y		CAC	GGG G1y	260	TCG	61	gcc Ala		ACT
	TCC		GGT Gly	460	GGT	510 ATG Met	u,	TAT		GCT Ala		GGA Gly
	CTC	410	ATG Met	4	AAA Lys	AAC		AAC Asn		GCC Ala	650	GGA Gly
360	AGC	•	GGT Gly		GAG Glu	ACA	550	CCA	600	\mathtt{TAT}	Φ.	GCT
	GAA Glu		ACT		ATC Ile	500 ATT Ile	55	GGC		TTT Phe		ATT Ile
	GGT	400	GGA G1y	450	CTC	GCC Ala		ATG Met		TGC Cys	0	ATG
350	GGC Gly	4	GTT Val		AAT Asn	TAT Tyr		CTG	290	TAC	640	CIC
•	CIC		CTA		CAG Gln	OCC CCC Pro	540	GGT	Ŋ	AAC Asn		GAC
	GAT Asp		ĠTG Val	440	GTT Val	490 ATT CC Ile Pr		TTG		TCC		GCT
340	TCC	390	GGA G1y	7,	$_{\rm GGG}^{\rm GGG}$	TTC	•	GAT	0	ACT Thr	630	GAG Glu
m	AAT Asn		GCT		GAC ASP	TTT Phe	30	ATC Ile	580	GCT		66C 61y

FIGURE 2

720	AGG Arg>	GAT Asp>		TTG Leu>		GGA Gly>	910	GAT Asp>	960	GGG G1y>	ACT Thr>
	CAA	CGT Arg		AGC	860	TTG	9	gcr Ala		GCT Ala	TCC
	TCT Ser	760 G GAC s Asp	810	GAG	w	TAT		AGG Arg		GAT Asp	O ACT Thr
710	TTA	£ \$		ATG Met		GAA Glu		CCA	950	GAA Glu	1000 GCG AC Ala Th
•	GCT	GAT Asp	•	GTT Val	850	GCA	006	GAT Asp	01	CTG	CAT His
	AGG	TGG Trd	800	TTG	8	ATT		ACT	:	AGT	GCT Ala
700	TGC	750 CCG Pro	w	GTA Val		ATT Ile		ATG Met	0	AGC	990 AAT Asn
7.	GCC	AGG Arg		GGA Gly		CCG	890	CAT His	940	GAG Glu	ATA Ile
	GTT Val	TCA	790	GCT	840	GCG	w	TAT Tyr		ATT Ile	TAC Tyr
	TTC Phe	740 GCC Ala	7.5	666 61y		GGA G1y		GCT		TGC	980 AAT Asn
069	GGA Gly	ACT		GAA		CGA Arg	0	GAT	930	TCT	GTC Val
	GGA Gly	CAG Gln		66C 61y	830	AAA Lys	880	TGT		TCC Ser	GAG Glu
	TTA	730 C CCT P Pro	780	ATG Met	w	ATG		AAT		GTC Val	970 CCT GAA Pro Glu
680	GGG G1y	G.A.		GTG Val		GCA Ala		GTC Val	920	${\tt GGT} \\ {\tt Gly}$	97 CCT Pro
•	ATT Ile	GAT Asp		TTT Phe	0	CAT His	870	GCA	O	CTT Leu	TCA
	CCA	AAT Asn	70	GGT Gly	4 820	GAA Glu		GGT Gly		$^{\rm GGG}_{\rm G1Y}$	GTC Val

FIGURE 2

	AAG Lys>		CAC His>	0.0	AAG GGA Lys Gly>	1200	GAG Glu>	GAA Glu>		TCA Ser>		GCA
,	TTC Phe	1100	GGA Gly	1150	-	-	CCC	CAT His		AAC Asn	1340	TCA AAT
1050	GTT Val	ä	ATC Ile		ATT Ile		AAT Asn	io CAA Gln	1290	CAC	Ħ	TCA
7	AAG Lys		ATG Met		ACA Thr	1190	TTC Phe	1240 CAG CAA Gln Gln	П	GGC		GGT
	AAG Lys	0.0	TCG	1140	ATT GCG	11	CAA Gln	AAG Lys		GGA Gly	0	
1040	ATC	1090	AAG Lys	(-)	ATT Ile		AAC Asn	230 AAC AAG Asn Lys	1280	TTC Phe	1330	TTA CTC
1	GCC		ACT		GCC	0	ATA Ile	1230 AAC Asn	12	GGA Gly		TGA
	AAT Asn		GCA Ala	1130	GAA Glu	1180	AGC	GCC Ala		TTC		CCA
30	ATA	1080	AAT Asn	7	CTT		CCC	GTT Val	0	TCA	1320	AAG Lys
1030	GAG		ATC		GGT Gly		CAT	1220 GAC ACA Asp Thr	1270	AAT Asn	П	TTC
	GCC		ACA	02	TCA GGG Ser Gly	1170	CTT Leu	12 GAC ASP		TCA		GCC
	CTT Leu	1070	ATC Ile	1120	TCA	1-1	TGG	TTC		ATC Ile	1310	TCA
1020	GAT	H	GAA Glu		GCA Ala		66C G1y	1210 GTG GAA Val Glu	1260	GCT Ala	4	TTC
	GGG G1y		AAG Lys		GGA Gly	1160	ACC	1210 GTG GZ Val G	(7)	GTT Val		GCT
	GCT Ala	09	ACC	1110	CTT Leu	11	ACC	TCA		AAT Asn	0	GTA Val
10	CTT	1060	AAC	• •	TGT		ATA Ile	CCA Pro	20	GTG Val	1300	GTT Val
						•						

TGURE 2

FIGURE 2 5/5

AATTTGTTGC TGAGACAGTG AGCTTCAACT TGCAGAGCAA TTTTTTACAT GCCTTGTCGT CGGAAGAGCG TAATACCGGG ATAGTTCCTT GATAGTTCAT TTAGGATGTT TTACTGCAAT AATCGAAGAT TATTTCCATT CTAATCCAGT CTCCGNCGAG TTTGAGAATC TATCTGTTTG TATTAGAAAG AACGAGGCAA GATTTTGTTT CATGTTTGTG TTTGTATTAC TTTCTTTTTG CCCTTGTCAA TGGCATTTAA GATAAGCTTA TAAAAAAAA AAAAAAAAA AAAACTCGAG GGGGGGCCCG GTACCCAATT CGCCCTATAG TGAGTCGTAT GACAATTCAC TGTCCGTCGG

	٠.		٠.									
09	ACTAAAGGGA ACAAAAGCTG GAGCTCCACC GCGGTGGCGG CCGCTCTAGA ACTAGTGGAT	120	GCTCAGGTGT	3 TGG r Trp		TCC		TCC		TGC	360	GGA Gly
	ACTA		3CTC.	r ACG s Thr		CGT	260	CTC	310	CCT	.,	TTC
20	AGA .	110	ICA (TGT CYS	210	CCA	5	ACT		GAT		CTC
	CTCT	••	GGTCGGCTCA	160 r TTC > Phe	•	GAC		AGG Arg		CTC	0	TCC
	CCG			CCT		AAC Asn		CGG Arg	300	TGC	350	GCT TCC Ala Ser
40	3008	100	TTCTTACTTG	3 TCC	200	GAC	250	CGC	(*)	CAA Gln		TTC Phe
	GGTG(CTTA(150 r GCG 1 Ala	7	TCC		CGT Arg		TTC Phe		GGA Gly
0	ပွဲ ပ	0		1 3 GTT E Val		TCA		TCC	290	ACC Thr	340	AAC Asn
30	CCAC	90	GGCACGAGTT	C ATG s Met		ACT	240	CTC	2	TCC		GAT Asp
	AGCT		GCAC	140 TCT TGC Ser Cys	190	CCC	••	CGC		GGA G1y		GGG G1y
. 20	TG G	80		14 T TCT a Ser		ATG		CTC		CGC	330	CTC
٠	AAGC		GCAGGAATTC	C GCT r Ala		TGC	230	CGG Arg	280	CTC	(*)	TTC Phe
•	ACAA		GCAG	o G ACC a Thr	180	GCA Ala		AAG Lys		TCC		CGC
10	GGA	70		130 3 GCG E Ala	,	GCT		CAC His		TGC	0	CAA
	AAAG		CCCCGGGGCT	A ATG Met		GTA Val		TCC	270	CAT His	32(CAG Gln
	ACT	٠	CCC	TCCA	170	CTC	220	CTT	N	TCC		AAC Asn

FIGURE 3 1/6

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ACT		GAA Glu		GTG Val		TAC	009	AAC Asn	TCT Ser		GAC
CGC Arg	٠	CAG Gln	200	GTT Val	550	GTT Val		GAG Glu	AAG Lys		ATG Met
66C 61y	450	GCA Ala	2	GTA Val		GAT Asp		ATA Ile	ATC	069	AGG Arg
400 CTC Leu	•	CCT		CGA Arg		CCC	290	GAG Glu	640 GAG Glu	Ψ	GAG AGG Glu Arg
AGG		CAA Gln		CAA AGG Gln Arg	540	GAC	55	AGT	GGA Gly		TCC Ser
CTG	440	ATG Met	490		٠,	CAT His		ATA Ile	GCC Ala	680	TTC
390 CAC His	4	GCT		AAG Lys	,	GGC Gly		GGC Gly	630 AGA ATT Arg Ile	99	AAG Lys
660		GTG Val		ACC	30	CTA	580	AGT	AGA Arg		CCA
CGC		GCT	480	GCT	53	CCT		ATA Ile	ACG Thr		GCC Ala
380 TCA AAT Ser Asn	430	ATG Met	•	CCT		ACT		GGA Gly	620 T CCC	670	GTG Val
TCA		GTC		AAA Lys		GTG Val	570	GAC	62 TTT Phe		TGG
CGT		GAG Glu	470	AAG Lys	520	GTG Val	u,	CTA	CAG Gln		66C 61y
CTT Leu	420	GGG G1y	4,	AAT Asn		GGC Gly		CTC	TCT Ser	* 099	GAT Asp
370 CCT Pro	•	TCC		ACA Thr		ATG Met	0.5	AAT Asn	610 TGC Cys	w	ACA
AAG Lys		CAT His		TCC	510	GGT Gly	560	AAC Asn	GAC Asp		TCC
TCC	410	TCC	460	GTC	u ;	ACA		TAC	TTC	650	TTT Phe

					•							
•	GAT Asp		TGT	840	GAT	TGT Cys		GAC		ACA		GAA
740	GCA	790	AAG Lys	w	AGC	TTT Phe		ATG	980	GCA Ala	1030	GGC
	TTA Leu		aga Arg		TTC	CCC	930	GCA	8	TGT Cys	77	AAA Lys
	GCA Ala		AAA Lys	830	GTA Val	880 AGT Ser	0,1	CTT Leu		GCC Ala		ATC Ile
730	aaa Lys	780	AAT	æ	AAG Lys	ATC		ATT Ile		ACT Thr	1020	ATA Ile
•	AAG Lys		CTC		ATG Met	AAG Lys	920	GCT	970	TCA	10	CAC
	GGC Gly		GAG Glu		GGT	870 AAG Lys	92	TCC		ATA Ile		AAC Asn
720	GCA	770	AAA Lys	820	GGC	8 TAT TYT		GGA Gly		TCG	0.	GCG Ala
7:	ACT	7.	ATG		TTG Leu	TCA Ser		ATG Met	096	TAT	1010	GCT
	CTG		GCG		GGA G1y	50 ACT Thr	910	AAT Asn	Oi	AAC Asn		AAT Asn
0	* ATG Met		GAT	810	TCC	860 AGG AC		ACA Thr		CCT		CTG
710	TAC	760	GAA Glu	w .	GGC Gly	CTG		ACC	950	GGC Gly	1000	ATA Ile
	CTT Leu		ACT		ATT Ile	GCT	006	TCT	9	ATG Met	П	TGT
	ATG		ATC Ile	800	CTC Leu	850 GAA Glu	0,	$ ext{TTT}$		TGG Trp		TTC Phe
700	TTC	750	GGA Gly	8	GTT Val	ATT Ile		CCT		GGA Gly	066	AAC
• '	AAG Lys	1-	GGT		GGA Gly	TCC	890	GTA Val	940	TTG	on.	AGT

FIGURE 3 3 OF 6

1080	GTT Val	AAT Asn		TTT Phe		CAT		AGT	1320	GCT Ala	TCG
ĭ	CCT	AAT Asn		GGA Gly	0	GAG Glu	1270	GGG	13	GGA Gly	GTC Val
	TTA	AGG Arg	1170	GAT Asp	1220	TTA	77	GGT Gly		GAA Glu	GGA G1y
20	GTT Val	1120 CAG Gln	H	CGT Arg		GAG Glu		CTA	0	CCT	360 TCC Ser
1070	GCC	TCA Ser		AAT Asn		GAG Glu	1260	TTT Phe	1310	CAC His	CAG Gln
	GCG	TTG	20	AGT	1210	CTT	77	GAA Glu		CCT	GCT Ala
	GAT	1110 CGA GCT Arg Ala	1160	GAC	,	CTT		GCG		GAG Glu	1350 GCC TTG Ala Leu
1060	TCG			TGG		TTA	0.0	\mathtt{TAT}	1300	ACC	13 GCC Ala
• •	66C 61y	TGC		CCA	1200	GTT Val	1250	ATT	-	ATG	AAG Lys
	GGT Gly	1100 GTA GCA Val Ala	1150	aga Arg	7	GGA Gly		ACC		CAC	1340 ATA GAG Ile Glu
1050	TGT Cys		• •	TCG		GCT		GCA	1290	TAC	1340 ATA G
ਜ	CTT	TTC		GCT	06	GAA GGA Glu Gly	1240	GGT Gly	12	GCC	TGC Cys
	ATG	GGT Gly	1140	AAA Lys	1190		F-1	AGA Arg		GAC	CTC Leu
40	* ATG Met	1090 GGA G1Y	H	ACC	٠	GGA Gly		AAA Lys	00	TGC	1330 3 ATC Ile
1040	GAC	TTG Leu		CCT		ATG Met	1230	AAG Lys	1280	ACT	GTG Val
	GCA	GGT G1Y	1130	GAC	1180	GTG Val	12	GCA		TTC	GGT

FIGURE 3 4 OF 6

	GCT		AAC Asn		CTT	1560	AGG	66C 61y		GTC		TCC
	CCT	20	CAA	1510	CTT	15	ATA Ile	GAA Glu		AAG Lys	0	TCA
1410	ACT	1460	GGC		CAC		GCA	GAC Asp	1650	AAA CTG Lys Leu	1700	AAC Asn
તે	TCC		TTC Phe		GGT	0	CAG Gln	1600 CCG	. 16	AAA Lys		CAT His
	ACT		TGT	1500	ATG ATC Met Ile	1550	GTT Val	GAC		GAG Glu		GGC G1y
00	GCA	1450	CAC His	1	ATG Met		GTA Val	GAA Glu	0	AAG Lys	1690	66C 61y
1400	CAT His	• •	GCC Ala		rcg		GCA Ala	1590 AAT TTG Asn Leu	1640	AAG Lys		TTC
	GCG Ala		CTC	0	ACC AAA Thr Lys	1540	GTT Val	15 AAT Asn		CCT		666 G1y
	ATA AAT Ile Asn	1440	GCT	1490	ACC	V-1	GCA Ala	ATT Ile		GGC Gly	1680	TTT Phe
1390	ATA Ile	ਜ	CAA Gln		TCC		GAA Glu	1580 CCA AAT Pro Asn	1630	GTC Val	7	TCA TTT Ser Phe
•	TAC		TAC		AGA GTG AAT Arg Val Asn	1530	GTA Val	1580 CCA A Pro A		CTC		AAT Asn
	AAT Asn	30	GAA Glu	1480	GTG Val	끔	GGC	CAT His		CTG	0	TCC
1380	GTA Val	1430	AAG Lys	• •	AGA		GGT Gly	ATC Ile	1620	AAA Lys	1670	TTG TCC Leu Ser
∺	GAC ASP		ATC Ile		CTG	0	GCT Ala	570 TGG Trp	16	GCA		GGT
	GAA Glu		GAT Asp	1470	GAG Glu	1520	GGA Gly	GGA Gly		GAT		GTC Val
1370	AGG Arg	1420	GGA	1,	AGT		GGA Gly	ACA Thr	1610	GTG Val	1660	AAG

FIGURE 3 5 OF 6

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1710	1720	1730	1740	1750	1760	
ATA CTA TTT GCC Ile Leu Phe Ala	r GCC CCC TGC 1	GC AAC TAG A ys Asn ***	AAAGAGTCTG	FGGAAGCCGA	AAC TAG A AAAGAGTCTG TGGAAGCCGA GAGTCTTTGA Asn ***	
1770	1780	1790	1800	1810	1820	
GAACTCATGC	GAACTCATGC ACGTTAGTAG	CTTCTTATGC CTCTGAAACC GAGATAGACC GGCTACTCGA	CTCTGAAACC	GAGATAGACC	GGCTACTCGA	
1830	1840	1850	1860	1870	1880	
GGGGATGCCA	AAGATACTCC	GGGGATGCCA AAGATACTCC TTGCCGGTAT TGGTGTTAAG AGATCACTGC TTGTCCCTTT	TGGTGTTAAG	AGATCACTGC	TTGTCCCTTT	
1890	1900	1910	1920	1930	1940	
TATTTTTT	TTCTTTTGAG	TATTITCTIC TICTITIGAG AGCITIAACC GAGGIAGICG IATTITCGAG CITITCGAAI	GAGGTAGTCG	TATTTCGAG	CTTTTCGAAT	
1950	1960	1970	1980	1990	2000	
ACATGTTCGT		TATCGGATCA ATGTGTTTCT TCTAAGATCA TTTGTAATGC ATATTTTGAA	TCTAAGATCA	TTTGTAATGC	ATATTTGAA	
2010	2020	2030	2040			
AAACCACATC	TCAGTATGCA	AAACCACATC TCAGTATGCA AAATAAAAA AAAAAAAAAA	* AAAAAAAAA	4444		

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Range:
Sequence

09*	CTACACCTCC	120	GCTCAATCGA	180	CTGCACAGGA AGTTACCACA	GGA ATG Gly Met>	,	AAT AAT Asn Asn>	320	GAT TGT ASD CYS>	370	TCC ACA Ser Thr>
20	CTA	110	GAG G	170	GGA A	220 GTG ACT Val Thr	270	TAC	m	TTT Phe		TTC
	GCCATGACTA		ACCG		CACA(TTC		ACC		TCT Ser
			ACCACCGGCA GGCACCGGAG			GTT Val		GAT GTT Asp Val	310	GAG Glu	360	AAG Lys
40	TTCGAGCCCT	100	cgca	160	GCTCTGCAAC	GTA	260	GAT Asp	3,	ATA Ile		ATC
	CGAG(CACC		rcTG(210 CGA Arg	••	CCT		GAG Glu		GAG Glu
		0		0		CGG		GAC		ATA AGT Ile Ser	350	GGA G1y
30	CTG(90	rccg	150	CTGT	CAG	250	CAT	300		(*)	GCT
	сстсесстес		GCCCATCCGC		AATGGCTGTG	200 ATC AAA Ile Lys	23	66C 61y		66C 61y		ATT
20		80		140	3C A			CTA		AGT	340	AGA
•	тсасстстта		TCGGATCCAG	Ĥ	CCGGGGAGGC	AGT		CCT	290	ACG Thr	36	ACG
	TCAC		rcgg,		2000	190 AAG CCA Lys Pro	240	ACT		GGA Gly		CCT
10	AGG 7	70		130) HHZ			GTG Val		GAT ASD		TTT Phe
	CGGCACGAGG		GCATCCTTGT	. •	GCTTCCCCTT	AAG Lys		GTG Val	280	CTT Leu	330	CAA
	CGG		GCA		GCT	AAG Lys	230	GGT Gly	7	CTG		GCT

IGURE 4

420	ATG Met>	ATC Ile>		CTC Leu>		GAA Glu>	610	TTC Phe>	099	TGG Trp>	TTT Phe>
· <u>.</u>	TTC Phe	GGA Gly		GTT Val	260	ATT Ile	6	CCT		GGA Gly	AAC Asn
	AAG	460 rr ggr in gly	510	GGA G1y	υ,	GCC Ala		GTA		TTG	700 ACG AGT Thr Ser
410	GAC	A A		TGC		GAT Asp		TGT Cys	029	gac Asp	
•	ATG Met	ACA		AAA Lys	55.0	AAT Asn	009	TTT Phe	v	ATG Met	GCA
	AGG	TTA	200	aga Arg	5,	TTC		CCC		GCA Ala	TGT Cys
400	AAG Lys	450 GCA Ala	υ,	AAA Lys		GTA Val		AAT Asn	0	CTT Leu	690 GCT Ala
4	TCC	aaa Lys		GAT Asp		AAG Lys	290	ATG Met	640	ATG Met	ACT
	CTC	AAG Lys	0	CTA	540	ATG Met	u)	AAG Lys		GCT	TCT Ser
	AAG Lys	440 GGC Gly	49	GAG Glu		GGA G1y		AAG Lys		TCA	80 ATA Ile
390	CCG	GCC		AAA Lys		GGT Gly	0	TAT	630	GGA	o TCG Ser
	GTG GCC Val Ala	ACT		ATG Met	530	ATG Met	580	TCA		ATG Met	TAC
	GTG Val	430 G CTG	480	GTG Val	u,	gca Ala		ATT Ile		AAT	670 CCC AAC Pro Asn
380	TGG	43 ATG Met		GAT Asp		TCA		AGG Arg	620	ACA	67 CCC Pro
	GGT Gly	TAC		GAA Glu	0	GGC G1	570	CTA	Φ	ACC	GGC
	GAT	CTT	470	ACC	52(ATT Ile		GCC		GCT Ala	ATG Met

FIGURE 4 2/6

ATT Ile>
GTG Val
gga gly
GCT Ala
GGA Gly
GAT
CCT
CAC His
CCT
GAG Glu
ACC
ATG
CAC His
TAC
GCC
GAT

3/6 3/6

	•	•										
0	GAA GAC Glu Asp>	1140	ATC Ile>	TTA Leu>		GCC Ala>		TGG Trp>	0	ACC Thr>	1380	GGT Gly>
1090	GAA	•	GAT Asp	GAG Glu		GCA Ala	1280	ACT GGG TGG Thr Gly Trp	1330	GAT ACC ASP Thr	П	GTC GGT Val Gly
	AGG		GGA Gly	30 AAC Asn	1230	GGA G1y	17			GTG Val		ATT AAG Ile Lys
	TCT	1130	GCT	1180 AAC AAC Asn Asn	•	CTC		AGG		ggc	1370	ATT Ile
1080	GTC Val	H	CCA Pro	CAA Gln		CTT	0.0	GCA ATA Ala Ile	1320	GAA GGC Glu Gly	딤	AAC Asn
••	GGA Gly		ACT	GGC Gly	1220	CAC	1270	GCA Ala	-	gat Asp		CTG
	TCA	20	TCC	1170 TTC	ਜ	GGT Gly		CAG Gln		CCA	.0	AGA Arg
1070	CAG	1120	ACA Thr	TGT Cys		ATT Ile		GTT Val	1310	AAC Asn	1360	GAG
ਜ	GCT		GCC	CAC His	01	TCA ATG Ser Met	1260	TCA GTA GTT Ser Val Val	딤	gaa glu		AAG Lys
	TTG		CAT	1160 CTT ATC	1210	TCA		TCA		TTG		GGC CCT AAG Gly Pro Lys
9	GCT	1110	GCA Ala	1. CTT Leu		aaa Lys		GTT Val	00	AAT Asn	1350	CCT
1060	AAG Lys	•	AAT	GCT		ACC	1250	GAA GCA GTT Glu Ala Val	1300	ATT Ile	П	GGC G1y
	GAG Glu		ATA Ile	50 CAA Gln	1200	TCT	7			AAT Asn		GTG Val
	ATA Ile	1100	TAC	1150 TAC C TYF G	• •	AAT Asn		GTG Val		CCG	1340	CTC
1050	TGC	Ħ	AAT	GAG Glu		GTG Val	01	GGT Gly	1290	CAT His	13	TTG
, ,	CTC		GTA Val	AAA Lys	1190	AAA Lys	1240	GGT Gly	П	ATC Ile		AAA Lys

FIGURE 4

TTG	TCT Ser	1390 AAT Asn	ICA Ser	TTC	1400 GGG '	1400 GGG TTT Gly Phe	GGT G1y	1410 GGG CAC 1 Gly His 2	AC AA	AAC TCG TCC ATA Asn Ser Ser Ile	1420 TCC Ser	ATA Ile	CTC Leu	TTC Phe>	
1430		7	1440		• •	1450		1460	-	14	1470		1480		
GCC	CCT	TAC	AAC TAG Asn ***>	TAG ***>)))	GGCGTTT		CATGTGGA ATTCTACTCA ATCTATCAAA	ATT	CTACT	CA AT	CTA	rcaaa	4	
	1490	90		1500	۰.		1510		1520	0	15	1530		1540	
GCTC	3AAGT	TT 1	rgagg	ACTC	ic Ac	SCATG	TTGG	GCTGAAGTTT TGAGGACTCC AGCATGTTGG TAGCTCCTTA CGTCTCTAGA CATGCCCATG	CCTT	A CGT	CTCTA	GA C	CATGC	CCATG	
	1550	20		1560	o *		1570		1580	0	15	1590		1600	
AGT	AGTTTTGTGT		GGGA	GCTG	T AC	3TCGG	AAACC	CGGGAGCTGT AGTCGGAACC ATGACGGATT GAGTACTCAT	GGAT	T GAG	TACTC		3GCGA	GGCGACACAG	
	1610	10		1620	0+		1630		1640	0	16	1650		1660	
GAT	GATATACTCC		TTGCTAGAAT	AGAA		FTAG	TGTTAGAGCA	CTATTCATTA	CATT	A TCC	тсссатттт		PTTCT	TTTCTGAAAT	
	1670	20		1680	o *		1690		1700	0	17	1710		1720	
CTCC	CTCC	rt A	CGGT	AGTT	G TZ	ACTTT	CGAG	CTCCCTCCTT ACGGTAGTTG TACTTTCGAG CGTTTCATCG AGTCAGTGAA GAAGAGAACA	CATC	G AGT	CAGTG	AA G	aaaga	GAACA	
	1730	30		1740	0 *		1750		1760	0	17	1770		1780	
AAGC	AAGCTAACTC		GGGCACGTAG	CGTA		ACCA	TAACCATTTG	CCCTTTGTTT	TGTT		тестстстат		TTAT	TTTATCGCCG	
	1790	06		1800	0 +		1810		1820	0	18	1830		1840	
TTTT	TTTTGTGGGT	T TE	TAAAATTTGT	TTTG		AACT	AGAC	AAAACTAGAC GACTGGTTTG	GTTT		TTTTCTCTTG	TG A	TCAT	ATCATTGGAG	

FIGURE 4 5/6

1900

1890

1880

1870

1860

1850

ATGTATGGCC ATATTTGCCT TTCATTGATG ATAAAAAAA AAAAAAAA AAAAAAAA

1920

1910

*
AAAAAAAAAAAA A

FIGURE 6/6

09	120	169	217	265	313	361	409	457	505
CTGGTACGCC TGCAGGTACC GGTCCGGAAT TCCCGGGTCG ACCCACGCGT CCGTCTTCCC	ACTCCGATCG TTCTTCTTCC ACCGCATCTC TTCTCTTCTC	CGCCGCC ATG CAT TCC CTC CAG TCA CCC TCC CTT CGG GCC TCC CCG CTC Met His Ser Leu Gln Ser Pro Ser Leu Arg Ala Ser Pro Leu 1	GAC CCC TTC CGC CCC AAA TCA TCC ACC GTC CGC CCC CTC CAC CGA GCA Asp Pro Phe Arg Pro Lys Ser Ser Thr Val Arg Pro Leu His Arg Ala 15	TCA ATT CCC AAC GTC CGG GCC GCT TCC CCC ACC GTC TCC GCT CCC AAG Ser Ile Pro Asn Val Arg Ala Ala Ser Pro Thr Val Ser Ala Pro Lys 35	CGC GAG ACC GAC CCC AAG AAG CGC GTC GTG ATC ACC GGA ATG GGC CTT Arg Glu Thr Asp Pro Lys Lys Arg Val Val Ile Thr Gly Met Gly Leu 50	GTC TCC GTT TTC GGC TCC GAC GTC GAT GCG TAC TAC GAC AAG CTC CTG Val Ser Val Phe Gly Ser Asp Val Asp Ala Tyr Tyr Asp Lys Leu Leu 65	TCA GGC GAG AGC GGG ATC GGC CCA ATC GAC CGC TTC GAC GCC TCC AAG Ser Gly Glu Ser Gly Ile Gly Pro Ile Asp Arg Phe Asp Ala Ser Lys 80	TTC CCC ACC AGG TTC GGC GGC CAG ATT CGT GGC TTC AAC TCC ATG GGA Phe Pro Thr Arg Phe Gly Gly Gln Ile Arg Gly Phe Asn Ser Met Gly 95	TAC ATT GAC GGC AAA AAC GAC AGG CGG CTT GAT GAT TGC CTT CGC TAC Tyr lle Asp Gly Lys Asn Asp Arg Arg Leu Asp Asp Cys Leu Arg Tyr 115

IGURE 5

553	601	649	697	745	793	841	688
GCC	666 61y	CTT Leu	GCC Ala 190	ATG Met	TGC Cys	ATG Met	66C G1Y
GGT G1y	GTT Val	TCT	TAT Tyr	CTG Leu 205	TAC Tyr	CTT Leu	GGA Gly
CTC Leu 140	CTG	CAA Gln	CCC Pro	GGT Gly	AAC Asn 220	gat Asp	TTG Leu
GAT Asp	GTG Val 155	GTT Val	ATC Ile	CTC	TCC	GCT Ala 235	666 G1y
GCC	GGA Gly	GGG G1y 170	TTC Phe	GAA Glu	ACT Thr	GAG Glu	ATT 11e 250
GAC	GCC Ala	GAC	TTC Phe 185	ATT Ile	GCC Ala	GGT Gly	CCA
GAG Glu	aga Arg	TCT Ser	CCT	GCT Ala 200	TGT Cys	CGT Arg	ATT Ile
CTT Leu 135	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala 215	CGC Arg	ATC Ile
TCT Ser	AAG Lys 150	GTC Val	ATC Ile	CTG	ACT Thr	ATC 11e 230	GCA Ala
AAG Lys	GAC Asp	ACT Thr 165	AAA Lys	GCC	TCC	CAT His	GCC Ala 245
AAG Lys	ATC Ile	CTG Leu	CGG Arg 180	TCT	ATT Ile	AAT Asn	GAG Glu
666 61y	AAG Lys	GGT Gly	CAC His	666 617 195	TCA	GCT Ala	ACT
GCC Ala 130	TCC	GGT Gly	GGT	ATG Met	TAT TYT 210	GCT Ala	660 617
GTC Val	CTC Leu 145	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala 225	GGA Gly
ATT Ile	CGC	GGA G1Y 160	GAG Glu	ACA Thr	CCA	CAT His	GCT Ala 240
TGC Cys	GAC ASD	ACA Thr	ATC Ile 175	ATT Ile	66C 61y	TTC Phe	ATT Ile

FIGURE 5

937	985	1033	1081	1129	1176	1224	1272
ACT Thr 270	GAA Glu	CGA Arg	gat Asp	TCT Ser	GTC Val 350	GCC Ala	aaa Lys
CAG Gln	GGT G1Y 285	aaa Lys	TGT Cys	TCC	GAG Glu	CTC Leu 365	ATC Ile
CCT Pro	ATG Met	ATG Met 300	AAC Asn	GTC Val	GAA Glu	gat Asp	GAT ASP 380
gac Asp	GTG Val	GCA Ala	ATC Ile 315	GGT Gly	CCT	666 61y	AAG Lys
gat Asp	TTT Phe	CAT His	GCA Ala	CTC Leu 330	TCA	GCT Ala	ACA Thr
AAC Asn 265	GGT Gly	GAA Glu	GGT G1y	GGT Gly	GTC Val 345	CTA	AAC Asn
AGG Arg	GAT ASP 280	TTG	GGA G1y	gat Asp	GGC Gly	ACT Thr 360	AAG Lys
CAA Gln	CGT Arg	AGC Ser 295	TTG	GCT Ala	GCT	TCT Ser	TTC Phe 375
TCT	GAC	GAG Glu	TAT TYY 310	AGG Arg	GAT	ACT	Grr Val
CTG	AAA Lys	CTG	GAG Glu	CCA Pro 325	GAA Glu	GCG Ala	AAG Lys
GCT Ala 260	gat Asp	GTG Val	GCA Ala	GAC Asp	CTT Leu 340	CAT	AAG Lys
AGG	TGG Trp 275	TTG	ATT Ile	ACT Thr	AGC Ser	GCT Ala 355	ATC Ile
TGC Cys	CCC	GTG Val 290	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala 370
GCT Ala	AGG Arg	GGA Gly	CCT Pro 305	CAC His	GAG Glu	ATA Ile	AAT Asn
GTG Val	TCT Ser	GCT Ala	GCA Ala	TAT TYr 320	ATT Ile	TAC	ATA Ile
TTT Phe 255	GCC	GGT Gly	gga gly	GCT Ala	TGC Cys 335	AAT Asn	GAG Glu

FIGURE 5 3/4

1320	1368	16	64	12	თ ა	62	68	7
13	13	1416	1464	1512	1569	1629	1689	1712
GGA GCC TCT GGA Gly Ala Ser Gly 395	ACC GGC TGG CTT Thr Gly Trp Leu	GTG GAG TTC GAC Val Glu Phe Asp 430	GTT GCG ATC TCG Val Ala Ile Ser 445	CT TTC TCG GCT la Phe Ser Ala 460	SAGTA CGGTTGTTCG	TCAAACCCAT TTAGGATACT GTTCTATGTA AAAAAAGTA AGGATTATCA CTTTCCCTTC	TAAAAAAAA AAAAAAGGGC	
CTT Leu	AAC Asn 410	TCC	AAT Asn	GTG GCT Val Ala	ATTGA	AGGAT	TAAAA	
C TGT s Cys	ATA Ile	G CCA u Pro 425	GTT Val	TCA GTC Ser Val	TGA TTACC CATTTCACAA GGCACTTGTC ATTGAGAGTA	AAGTA	ATATTTATT	
CAC His	GGA Gly	GAG	GAA Glu 440	Se	CAC	AAA	ATT	
GGA	AAG Lys	CCT	CAC His	AAC Asn 455	္က်ပ္သ	A.	AT	
ATC Ile 390	ATT Ile	AAT Asn	CAA Gln	CAC His	CACAA	ATGTA	VAATT	
ATG Met	ACT Thr 405	TTC Phe	AAG CAG (Lys Gln (GGC	ATTTC	rtctz	GAATGAAATT	E
TCA	GCG Ala	CAA Gln 420	aag Lys	GGA Gly	ស ស	F.	₹	A GCT
AAG Lys	ATA Ile	AAT Asn	AAG Lys 435	TTC Phe	TTAC	BATAC	TCCAGTTTGA	GGCCGCTCTA GAGGATCCAA
ACT Thr	GCT	ATT Ile	AAC	GGA G1Y 450		rtago	CCAG	3AGG?
GCA Ala 385	GAA Glu	AGC	GCC Ala	TTT Phe	CCA Pro 465	PAT 1		TA G
AAT Asn	CTT Leu 400	CCC	GTT Val	TCA	AAG Lys	ACCC	таатсствтс	GCTC
ATT Ile	GGT Gly	CAT His 415	ACT	AAT Asn	TTC	TCA	TAAT	၁၁၅၅

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Range:
Sequence

0 *	ည္ဟ	TCC		•						•	4
	TCC	10 CCT Pro	,	TCC	210	CGT Arg		CGG Arg		GTC Val	CTA Leu
	TTATCTCCGC	110 TCC CC Ser P1	160	TCC		ATC Ile		AAG Lys		GAC	350 ATC AGC Ile Ser
20		CAC 7		CCC		GTC Val		AAG Lys	300	TCC	
	נפככי	CTC (Leu 1		TCC	200	CCC	250	CCC AAG Pro Lys	.,	GGC	GGC G1y
	TCT	100 TCC (Ser 1	150	AAT Asn	2(CTC		GAC		TTC Phe	AGC Ser
40	ATT	CAA 7 Gln 8	***	CTC		AGC		TCC	0	GTC Val	340 GAG Glu
	TTT	ATG C		CGC		GCC	240	GAG Glu	290	TCC	66C 61y
_	AC.	Ö	o	TTC	190	CGC	N	CGC		GTC	TCC
30	GGTCGACCCA CGCGTCCGGG CTTTCCGACC ACATTTCATT TCTTGCCTCG	ລອລວອລລອລວ 06	140	CCC		CGT Arg	,	AAG Lys		CTC	30 CTC Leu
	TTCC	ອວວອເ		GAG Glu		CTC	0	CCC	280	GGC	3 CTG Leu
20	ຼິວ			CTC	180	CCC Pro	230	GCC		ATG	AAG Lys
(1)	SCCCC	80 CCGTCGTTCG	130	CCT	н	CGC		TCC		GGC	
	GCGI	CGTC		TCC		CTC		GCC	270	ACC	320 TAC GAC Tyr Asp
10	CA			CCC	0	GCT	220	ACC	73	ATC	TAC
	GACC	70 CGCTCCTCCG	120	CGC	170	GCC		GCC		GTC	GCC
	GGTC	CGCT	Н	CTC		GCC Ala		GCT Ala	260	GTC	310 GAC ASP

	CAG Gln	150	CGG		GCT		AAG Lys	GTC Val		ATC Ile	06	CTG	
400	GGC G1y	7	GAC		AAG Lys		GAT Asp	ACT Thr	640	AAG Lys	9	GCG	
	GCC		AAC		AAG Lys	540	ATT	590 CTA AC		CGG Arg		TCT	
	TTC	440	AAG Lys	490	GGC	۵,	AAG Lys	66C 61y		CAC His	089	GGG	
390	AGG	4,	GGC		GCC		TCC	GGT Gly	630	$_{\rm GGT}^{\rm GGT}$	39	ATG Met	
•	ACC		GAC		GTC Val	30	CTC	580 ATG Met	· .	AAA Lys		AAC Asn	
	CCC		ATC Ile	480	ATT Ile	53	TCC	GGT Gly		GAG Glu		ACA Thr	φ
380	TTC	430	TAC		TGC		CAA Gln	ACC	620	ATC Ile	670	ATT Ile	FIGURE 2/5
ñ	AAA Lys		GGC		TAC	•	GGC Gly	570 GGA Gly		CTC		GCC Ala	FIG 2
	TCC		ACG	470	CGC Arg	520	GCC Ala	GTT Val		AAT Asn		TAT Tyr	
	GCT	420	GCG	4	CTC		CTC	CTA		CAG Gln	660	CCA	
370	GAC	•	AAC Asn		TGC		gat Asp	50 Grg Val	610	GTT Val	v	ATT Ile	
	TTC		TTC		gat Asp	510	GCC Ala	560 C GGA GT a Gly Va		666		TTC	
	CGC	410	66C G1y	460	gac Asp	.,	GA	GC Al		GAC	0.0	TTT Phe	
360	GAC	4	CGT Arg		CTC		GAA Glu	AGG Arg	009	TCT	65	CCG	
•	ATC Ile		ATC		CGG	200	CTC	550 GAG Glu	v	TTC		TCC	
					•								•

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	ACT		ATC Ile	GCG		TCT	930	GAC		GAG Glu		TAT Tyr
	TCA		CAT His	30 GCT Ala	880	TTA	O1	AAG Lys		ATG Met		GAA Glu
	ATT Ile	780	AAT Asn	830 GAG GC		GCT		gat Asp		GTT Val	1020	GCA Ala
730	TCG	•	GCC	ACT		AGG Arg	920.	TGG	970	TTG	1	ATT
	TAT		GCC	GGA Gly	870	TGC	6	CCG		GTA Val		ATT Ile
	AAC	022	GCT	820 GGA G1y	w	GCC Ala		AGG Arg		GGA Gly	9	CCG
720	CCA	7.2	TAT Tyr	GCT Ala		GTT Val	i	TCA	096	GCT Ala	1010	GCG Ala
	GGC		TTT Phe	ATT Ile	860	TTC Phe	910	GCC	0,	GGG G 1y		GGA Gly
	ATG Met		TGC	810 ATG Met	8	GGA G1y		ACT		GAA		CGG Arg
710	CTG	760	TAC	CTG		GGA G1y		CAG Gln	950	$_{\tt G1Y}^{\tt GGT}$	1000	AAA Lys
7	GGT		AAC Asn	GAC Asp		TTA	006	CCT Pro		ATG Met	•	ATG
	TTG		TCC)0 GCT Ala	850	GGT Gly	01	GAT Asp		GTG Val		GCA
	GAT Asp	750	ACT	800 T GAG GC' Y Glu Ala		ATT Ile		GAT Asp		TTT Phe	066	CAT His
700	ATC Ile		GCT	9 5		CCA	890	AAT Asn	940	GGC G1y		GAG Glu
	GCC		TGT	CGA Arg	840	ATT Ile	8	AGG		GAT Asp		TTG Leu
	CTT	740	GCA Ala	790 CGC Arg		GTC Val		CAA Gln		CGT Arg	980	AGC

FIGURE 6 3/5

AGG Arg		GAT Asp	1170	ACT Thr		GTT Val		ATC Ile	ATT		AAT Asn
GAT CCA	1120	GAA	ਜ	GCG		AAA Lys		ATG Met	1310 GCA ACC Ala Thr	1360	CAA TTT Gln Phe
10 GAT ASP	•	CTC		CAT His		GCC ATT AAG Ala Ile Lys	1260	AAG TCA I	1310 GCA AC Ala Ti	••	
. Pr		AGT	0.0	GCT	1210	ATT Ile	11	AAG Lys	ATC Ile		AAT Asn
ATG	1110	AGC	1160	AAT GCT Asn Ala	``	GCC		ACT	GCC	1350	ATT Ile
1060 TAT CAT ATG A	H	GAG		ATA		GAG ATA AAT Glu ile Asn	: 09	AAT GCA A	1300 CTT GAA (Leu Glu	ä	CCC AGC ATT AAT Pro Ser Ile Asn
TAT		ATT Ile		TAC	1200	ATA Ile	1250	AAT Asn	CTT		CCC
55	00	TGC	1150	AAT Asn	Ä	GAG Glu		ATC Ile	GGT Gly	0.1	CAT
OSO GAT ASD	1100	TCG	••	GTC		GCC		AAA Lys	1290 A GGA E Gly	1340	CTT
1050 C AAC TGT GAT G 1 Asn Cys Asp A		TCC		GAG	90	GAT CTT Asp Leu	1240	GAA ATC 1 Glu Ile 1	11 TCA Ser		TGG
AAC Asn		GTC	1140	GAA Glu	1190	GAT Asp	П	GAA Glu	GCA		GGC Gly
E	1090	GGT GTC Gly Val	H	CCT		GGG		ACC AAG Thr Lys	1280 CTT GGA Leu Gly	1330	ATA ACC ACC Ile Thr Thr
1040 GCA G1 Ala Ve	• •	CTT Leu		TCA		GCT	1230	ACC Thr	1280 CTT GG Leu G1		ACC Thr
GGT Gly		$_{\rm GGG}$	0 0	GTC Val	1180	CTT C	13	AAC	TGT		ATA Ile
GGA Gly	1080	GAT Asp	1130	GGG G1y		ACT Thr		AAG Lys	CAC His	1320	GGA Gly
1030 TTG Leu	1(GCT		GCC		TCT Ser	1220	TTC	1270 GGA Gly	13	AAG Lys

GURE 6

1410	Gag caa Gln Gln		GGG CAC	1510	ATTCT ACTTGGTTCA	1570	TAAATGCCTT	1630	AGCCATTTAG	1690	CTCTGATTTA	1750	GTTATTTAAG		CT
1400	AAC AAA AAG Asn Lys Lys	1450	GGA TTT GGA Gly Phe Gly	1500	TGA * * *	1560	AAATGCACAC CAGTTGCTGA GATAGGGCTT CAACTTGCAG AGCAATTTTT TAAATGCCTT	1620	GTCCTTTGAT AGTTCCTCGA AGCCATTTAG	1680	TAAATCTAGT	1740	TGTTGTCAAT GTTATTAAG	1800	ATCCAGCTTA
1390	ACT GTT GCC Thr Val Ala	1440	AAT TCT TTT Asn Ser Phe	1490	TTC AAG CCA Phe Lys Pro	1550	CAACTTGCAG	1610	GTCCTTTGAT	1670	ATTCCCATTT	1730	GTCATGTTTG	1790	GCTCTAGAGG
	GAC TTC AAC ASP ASP Phe Asn 1	1430	GCT ATC TCG A	1480	TTC TCA GCT I	1540	GATAGGGCTT	1600	CGTAATACCG GAATAGGTCG	1660	TACTGTAATA ATCGAAGATG	1720		1780	AAGGGCGGCC
1380	CCA TCG GTG GAPro Ser Val As		AAC GTC Asn Val	1470	GTG GCA Val Ala	1530	CAGTTGCTGA	1590		1650		1710	TGTATTAGAA AGACCAATGA AAGATTTTGT	1770	ATAAAGCAAA AAAAAAAAA AAGGGCGGCC
1370	CCC GAG CC. Pro Glu Pro	1420	CAT GAA GTG His Glu Val	1460	AAC TCG GTT Asn Ser Val	1520	AAATGCACAC	1580	GTCGGAAGAG	1640	GATGATGTTT	1700	TGTATTAGAA	1760	ATAAAGCAAA

FIGURE 6 5/5

Sequence Range: 1 to 2369

						•						
	CACGCGTCCG CATAAAAGAG	120	CTTCGATTCA TTACCATACC	180	ATTCCGCTGA TCCATTTTCC GCCTTTTCCG GGTCTTTCAT CCCAAAGGGT ATCCTTTTCT	r TCC r Ser>	280	G TCT	330	r CCT r Pro>		A CTA
	CAT		TTA(ATC	230 GCC TCT Ala Ser	•	ATG Met		TCT Ser		CCA
20	SCG	110	ICA	170	GGT	GCC Ala		TGC		ATC TCC Ile Ser	370	GCC
	SCGT		GAT		AAAG	GCC		GCC Ala	320	ATC Ile	'n	TGC
	CAC				က	220 G CCT it Pro	270	GCC	(*).	TCC		CAA
40	BACC	100	rcat	160	rcat	22 ATG Met		CTT		CCT		TCC
	CGGGTCGACC		CTCCTTTCAT		CTTJ	rcca		CTC	310	CCG	360	CTC TCC Leu Ser
_	SS			_	9	GG	260	TGG	Э.	CTT Leu		ATT Ile
30	ATTCC	90	CACC	150	PTCCG	210 AGTTC	N	ACG		CCT		CGG
	CCGGAATTCC		AGAGAGGG ATCCATCGAA TGCGGCCACC		CCTT	200 210 220 CTCAAAGGGT CAGTCAGTTC CCTCCA ATG CCT Met Pro		TGT Cys		GAC	350	CGC
20	E C	80	Y. T.	140	ပ္ပ	200 GGT C	250	CTC	300	TCC		CGC
•	ACCG(w	ATCG?	17	FTTT	20 1AGG0	2	CCT CTC Pro Leu		CCC TCC Pro Ser		TCC
	GTACGCCTGC AGGTACCGGT		ATCC/		ICCA.	CTCA		TCC		CAC	340	CGC CTC Arg Leu
10	ည်	70	366	130	rga :			GCT	290	TTC Phe	36	CGC
•	CCC		3AGA(• •	CGC	190 ATCCTATCTT	240	CTC	(4	Ser		CGA
	GTA(AGAC		ATT(ATCO		CTG		ACC		CGC

FIGURE 7

	GTC Val>	TCC Ser>	0	CGG Arg>	570	CTG Leu>		CAG Gln>		CAT His>	ATA Ile>
	CTC	470 ACA Thr	52(CAC His		GCT	. •	AAA Lys		GGC	710 GGC G1y
420	ACC Thr	TAT Tyr		AGG Arg		GTG Val	610	ATC Ile	660	CTA	AGT
	CAT His	TAC		CGC Arg	260	GCC	61	AGT Ser		CCT	ACG
	TTC	50 GAC ASP	510	ACC	υ,	ATG Met		CCA		ACT	700 T GGA P Gly
410	AGT	460 C CAT GA S His AS		ACC		GCA Ala		AAG Lys	650	GTG Val	7 GAT ASD
•	TC(Se)	TGC Cys		CGC Arg	550	GAG Glu	600	AAG Lys		GTG Val	CTT Leu
	GGA G1y	CCC	200	CCC ATT (Pro Ile)	ິນ	AGG		AAG Lys		GGT	CTG
400	CGC	450 GAG Glu	. ,	CCC		TCC		ACA Thr	640	ATG Met	690 AAT Asn
4	CTC	TTC		AGA Arg		CCT	290	ACC	9	GGA	AAT Asn
	GCC	TGC Cys	490	TCC Ser	540	TCC	۵,	GTT Val		ACT	TAC
	TCC	440 GCC Ala	4	GGA Gly		GCT		GAA Glu		GTG Val	680 IT GTT TTC 1 IP Val Phe 1
390	TCC	CTC Leu		TTC		CGA	280	CAG Gln	630	GTT Val	GTT Val
	GCT	TAC		TTG	530	AAT	28	GAA Glu		GTA	G A
	TCT	30 TCT Ser	480	TCC	υ,	CTC		CCT		CGA Arg	70 CCT Pro
380	CCT	430 ACC TC Thr Se		GCA		AGG		CAA	620	CGG Arg	670 GAC CC ASP PI

FIGURE 7

	•							•				
0	GCT Ala>	810	CTC Leu>		AAG Lys>		CTA Leu>	ATG Met>	0	AAG Lys>	1050	GCT Ala>
760	ATT Ile		AAG Lys		66C 61y		GAG Glu	950 GGA G1Y	1000	AAG Lys	-	TCA
	AGA Arg		CCG	850	GCT Ala	900	AAA Lys	GGT		TAT Tyr		GGA G1y
	ACG Thr	800	GCC Ala	8	ACC Thr		ATG Met	ATG Met		TCA	1040	ATG Met
750	CCT	w	GTG Val		CTG		GTG Val	940 TCA GCA Ser Ala	066	ATT Ile	7(AAT Asn
	TTT Phe		TGG Trp		ATG Met	890	GAT Asp			AGG Arg		ACA Thr
	CAA Gln	790	GGT Gly	840	TAC	~	GAA	66C 61y		CTA	30	GCT ACC Ala Thr
740	GCT Ala	7.	GAT		CTA		ACC Thr	ATT Ile	980	GCC Ala	1030	
•	TGT		ACA Thr		ATG	880	ATC Ile	930 CTC Leu	0,	GAA Glu		TTC
	GAT		TCC	830	TTC Phe	88	GGA Gly	GTT Val		ATT Ile		CCT
730	TTT Phe	780	TTC Phe		AAG Lys		GGT Gly	GGA Gly	970	GCC	1020	GTA Val
7.	ACC		TCT		GAC Asp		GAT Asp	920 TGC Cys	6	GAT Asp		TGT Cys
	GAG Glu		AAG	820	AGG ATG Arg Met	870	ACA Thr	aaa Lys		AAT Asn		rrr Phe
	ATA Ile	770	ATC Ile	8	AGG Arg		TTA Leu	AGA		TTC	1010	CCC
720	GAG Glu		GAG Glu		AAG Lys		GCA Ala	10 AAA Lys	960	GTA Val	7(AAT Asn
	AGC		GGA G1y		TCT	860	AAA	910 GAT AJ ASP L	,	AAG Lys		ATG

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FIGURE 7

			·									
	TCT Ser>		CAT His>	GCG Ala>	40	TTG Leu>	1290	AGT Ser>		CTA Leu>		GAA Glu>
	ATA Ile		AAC Asn	1190 12 GAT 21 ASP	1240	GCT		GAC		CTA Leu		GCA Ala
0	TCG	1140	GCG	11 TCA Ser		CGA Arg		TGG Trp	0	CTA	1380	TAC
1090	TAC	-	GCT	GGC Gly		TGC	1280	CCA	1330	GTG Val		ATT Ile
•	AAC Asn		AAT Asn	ე. ქ	1230	GCA	12	AAA GCT TCA AGA CCA Lys Ala Ser Arg Pro		GGA Gly		ACT
	CCC	1130	ATG Met	1180 TGC G(Cys G		GTT Val		TCA		GCT	1370	GCG Ala
1080	666 61y	11	ATA	CTT Leu		TTT Phe	0	GCT	1320	GAA GGA Glu Gly	H 3	$_{\rm GGT}^{\rm GGT}$
⊣	ATG Met		TGT	ATG Met	1220	GGT	1270	AAA Lys	-	GAA Glu		aga Arg
	TGG	0	TTT Phe	170 GTG Val	12	GGA Gly		ACT	•	666 61y	0	AAA Lys
1070	GGA Gly	1120	AAC Asn	1 GAT ASP		ATG Met		CCT	1310	ATG Met	1360	AAG Lys
10	TTG		AGT	GCA	0	GGT Gly	1260	GAC Asp	13	GTT Val		GCA
	GAC		ACG Thr	1160 3C GAA 3Y Glu	1210	ATT Ile		TCC		$ extbf{T}$ T $ extbf{P}$ Phe		CAT
0	ATG	1110	GCA	11 GGC GlY		CCT		AAT Asn	00	GGA Gly	1350	GAG Glu
1060	GCA	-	TGT	aga Arg		ATA Ile	1250	aga Arg	1300	GAT ASP	•	TTG
	CTT Leu		GCT	00 ATC Ile	1200	ATC Ile	12	CAG Gln		CGT		GAG Glu
	ATG	1100	ACT	1150 ATA ATC Ile Ile	П	GTA Val		TCC		AAT Asn	1340	GAG

SUBSTITUTE SHEET (RULE 26)

•											
CCT	0	GCT Ala>	1530	GCC Ala>		CAC His>		ATG Met>	GTA Val>	02	GAA Glu>
1430 CC GAG hr Glu	1480	TTG	-	CAT His		ATC Ile		TCA	1670 GTT TCA GTA Val Ser Val	1720	TTG
ACC Thr		GCT		GCC	. 02	CTT Leu	1620	AAA Lys	1 GTT Val		AAT Asn
ATG Met		AAG Lys	1520	AAT Asn	1570	GCT	•	ACC	GCA		ATT Ile
0 CAC His	1470	GAG Glu	15	ATA Ile		CAA Gln		TCA	50 GAA Glu	1710	AAT Asn
1420 TAC CZ TYF H	П	ATA		TAC		TAC Tyr	1610	AAT Asn	1660 GTG GZ Val GJ		CCG
GAT GCC Asp Ala		TGC Cys	0.	AAT Asn	1560	GAG Glu	ř	GTT Val	GGT		CAT His
GAT	1460	CTC	1510	GTA Val	-	AAA Lys		AAA Lys	GGT Gly	1700	TGG ATC Trp Ile
410 TGC Cys	14	ATT Ile		GAC		ATC	00	TTA	L650 GCC Ala	ਜ	
1410 ACT TGC (Thr Cys A		GTG Val		GAA Glu	1550	gat Asp	1600	GAG Glu	1650 . GCA GCC GGT G . Ala Ala Gly G		666 61y
TTC Phe	0	GGA G1y	1500	AGG Arg	Ħ	GGA		AGA Arg	GGA Gly	0	ACT Thr
1400 GAGT Y Ser	1450	GCT Ala	П	TCT		GCT Ala		AAC Asn	1640 CTT CTC Leu Leu	1690	AGG Arg
1400 GGG AGT Gly Ser		GGA Gly		GTC Val	0.	CCG	1590	CAA Gln	16 CTT Leu		ATA
GGT Gly		GAT Asp	1490	GGA Gly	1540	ACT	,	66C 61y	CAC His		GCA
O. CTA Leu	1440	CCT Pro	14	TCA		TCC		TTC	30 GGT Gly	1680	cAG Gln
1390 TTT CTA Phe Leu	Н	CAC		CAG Gln		ACA	1580	TGT Cys	1630 ATT GGT Ile Gly	~	GTT Val

FIGURE 7 5/7

FIGURE 7 6/7

	•														
1770	AAG AAG Lys Lys>	•	TTT GGT Phe Gly>	1870	GTTTCCGTGT	1930	GTTGGTAGCT	1990	GAACCATGAC	2050	GTAGAGCAAT	2110	GTTGTACTTT	2170	CACGTAGTAA
1760	GTG GGT CCT Val Gly Pro	1810	TCA TTT GGG Ser Phe Gly	1860	ATC TAG GAC Ile ***>	1920	ACTCCAGCAT	1980		2040		2100	CCTTGCAATA	2160	TTAACTCGGG
1750	AAA TTG CTC Lys Leu Leu	1800	TTG TCT AAT Leu Ser Asn	1850	GCC CCT TAC Ala Pro Tyr	1910	AGTTTTGAGG	1970	CCATGAGTTT TGTGTCCGGA GCTTTAGTCG	2030	ACTCCTTGCT AGAATTGTTG	2090	AAATCTCCCT	2150	AACAAAGCTG
	GGC GTG GAT ACA A Gly Val Asp Thr L	1790	GTC GGT Val Gly	1840	CTC TTC Leu Phe	1900	CAAAGCTGA	1960	CATGAGTTT	2020	CACTTGATAT	2080	TTTTTCTCTG	2140	rgaagaagag
1740	GAA Glu	1780	AAC GTT AAG Asn Val Lys	1830	TCG TCC ATA Ser Ser Ile	1890	GTGGAATTCT ACTCAACATA TCAAAGCTGA AGTTTTGAGG	1950	CTAGACATGC C	2010	CTCATGGCGA C	2070	TCATATTTT 1	2130	CGAGCTTTTC ATCGAGTCAG TGAAGAAGAG AACAAAGCTG
1730	AAC CCA GAT Asn Pro Asp	17	GAG AGA CTG Glu Arg Leu	820	GGG CAC AAC Gly His Asn	1880	GTGGAATTCT .	1940	CCTTACGTCT	2000	GGATTGAGTA	2060	ATTCATTATC	2120	CGAGCTTTTC

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				CTCTAGAGG	2360 AGGGCGCCG CTCTAGAGG
AAAAAAAAA	AAAAAAAA	AAAAAAAAA	AAAAAAAAA	TGGAAATAAA AAAAAAAA AAAAAAAA AAAAAAAA AAAAAA	TGGAAATAAA
2350	2340	2330	2320	2310	2300
ATGTATGTTT	TAATTGGGGR	TTCTCATTGA	TTGGTTTGTT	AACTAGAAGA CTGGTTTAGA TTGGTTTGTT TTCTCATTGA TAATTGGGGR ATGTATGTTT	AACTAGAAGA
2290	2280	2270	2260	2250	2240
AAATTTGTAA	TGTGGTTTTA	ATCACCGTTT	TCTCTATTTC	CCATITIGCCC TITGITITIGC TCTCTATITIC ATCACCGTIT IGIGGITITA AAATTIGTAA	CCATTTGCCC
000	0 *	0177	2200	2190	2180

FIGURE 7

Sequence Range: 1 to 2374

0 *	CACACCAAAC	120	ACAGACAGAC	180	TCTTCGATTC	240	TCCCAAAGGG	300	CCTGCCGCCT	360	TCTACCTCCT	420	CTCTCCCGCC	480	CGCGGATCCA
20	GACGCCAACC	110	AGACAGACAG	170	CCTCCTTTCA TCTTCGATTC	230	CGCCTTTTCC GGGTCTTTCA TCCCAAAGGG	290	TATCCTATCT TCTCAAAGGG TCAGTCAGTT CCCTCCAATG CCTGCCGCCT	350	CGCCTGCATG	410	TCGCCGACGC	470	ರಾದಂತಿಂದಾರ
40	ACGCGTCCGC	100	CATTGGCAGC	160	ATGCGGCCAC	220	CGCCTTTTCC	280	TCAGTCAGTT	340	GGCTCCTTGC	400	CCGCCTTCCA TCTCCTCTCC	460	CTTCTGCTTC
30	GGGTCGACCC	06	AGACGGACGC	150	GATCCATCGA	210	ATCCATTTTC	270	TCTCAAAGGG	330	CTCTGTACGT	390		450	GCCCCACTAC
20	-A-CNTGGTC CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC CACACCAAAC	80	TICCICAGCI ICICITCICA AGACGGACGC CATIGGCAGC AGACAGACAG ACAGACAGAC	140	CCATAAAAGA GAGAGAGG GATCCATCGA ATGCGGCCAC	200	CATTCCGCTG	260		320	CTTCCCTGCT CGCTTCCCCT CTCTGTACGT GGCTCCTTGC	380	CGACCCTCTT	440	GCCGGATTCT CTCCCAATGC GCCCCACTAC CTTCTGCTTC CTCCGCCCTC CGCGGATCCA
10	-A-CNTGGTC	70	TTCCTCAGCT	130	CCATAAAAGA	190	ATTACCATAC	250	TATCCTTTTC	310	CTTCCCTGCT	370	TCCACCCTC	430	GCCGGATTCT

FIGURE 8

540	GACTACTATA	009	CGGAGGCTCA	099	ACAGGAAGTT	720	AATGGGTGTG	780	ATGGAACGAG	840	TTGCTGGAGA	006	GGATGGACAA	096	GTTCATGCTA TACATGCTGA CTGCTGGCAA GAAAGCATTA ACAGATGGTG GAATCACCGA	1020	ATAAAAGAAA ATGCGGAGTT CTCATTGGCT CAGCAATGGG
530	GCCCTGCCAT	590	CCGCAGGCAC	650	TGCAACCTGA	710	TTGTGACTGG	770	AATCTGCTTG	830	CCTACGAGAA	890	CTCTCTAAGA	950	ACAGATGGTG	1010	CTCATTGGCT
520	CCTGCTTCGA	580	CTTGTTCGGA TCCAGACCCA TTCGCACCAC CCGCAGGCAC	640	GGAGGCAATG GCCGTGGCTC TGCAACCTGA ACAGGAAGTT	700	CGGCGAGTAG TTGTGACTGG	760	TTTCTACAAT AATCTGCTTG	820	TGCTCAATTT	880	GGCCCCGAAG	940	GAAAGCATTA	1000	ATGCGGAGTT
510	TCTTACCTCG	570	TCCAGACCCA	630	GGAGGCAATG	069	TATCAAACAG	750	ACCTGATGTT	810	CCTTTGATTG	870	ATGGTTGGGT	930	CTGCTGGCAA	066	
200	GITTCCATAC CCTCGTCACC TCTTACCTCG CCTGCTTCGA GCCCTGCCAT	560	CTTGTTCGGA	620	CCCTTCCAGG	680	ACCACAAAGA AGAAGCCAAG TATCAAACAG	740	TAGGCCATGA ACCTGATGTT	800	TGGCATAAGC GAGATAGAGA CCTTTGATTG TGCTCAATTT	860	GATCAAGTCT TTCTCCACAG ATGGTTGGGT GGCCCCGAAG	920	TACATGCTGA	980	AGATGTGATG AAAGAGCTAG
490	GTTTCCATAC	550	CATCCGCATC	610	ATCGAGCTTC	670	ACCACAAAGA	730	GTGACTCCTC	790	TGGCATAAGC	850	GATCAAGTCT	910	GTTCATGCTA	970	AGATGTGATG

FIGURE 8 2/5

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	AGAAGATGAA	1140	CAATGGACTI	1200	ACTTTTGTAI	1260	есееееест	1320	CTTTGTCCCA	1380	ATGGATTTGT	1440	AGAAAAGAG	1500	ACCACATGA
9	ATTTCATATA	1130	GCTATGCTTG CAATGGACTT	1190	GCAACGAGTA ACTTTTGTAT	1250	GTGATGCTTT	1310	GCATGCCGAG	1370	AGTAATCGTG	1430	GAGCATGCAA	1490	TGCGATGCCT
0007	AGCCCTAAGG	1120	TATGGGATCA	1180	тастесттет	1240	CGAAGCAGAT	1300	AGGTTTTGTT	1360	ACCATGGGAC	1420	TGCTACTACT AGAGGAGTTG GAGCATGCAA AGAAAAGAGG	1480	GAGTTTCACT
1050	ATGCCATTGA	1110	CTACCACAAA	1170	астссататс	1230	TAATCAGAGG	1290	TTGGTATGGG	1350	AAGCTTCAAG	1410		1470	TTCTAGGTGG
1040	TGGAATGAAG GTATTCAATG ATGCCATTGA AGCCCTAAGG ATTTCATATA AGAAGATGAA	1100	TCCCTTTTGT GTACCTTTCG CTACCACAAA TATGGGATCA	1160	GGGATGGATG GGGCCCAACT ACTCGATATC TACTGCTTGT	1220	GCGAACCATA TAATCAGAGG CGAAGCAGAT GTGATGCTTT GCGGGGGCTC	1280	AGATGCGGTA ATCATACCTA TTGGTATGGG AGGTTTTGTT	1340	GACCCTACTA AAGCTTCAAG ACCATGGGAC AGTAATCGTG	1400	GGAGCTGGAG	1460	TGCGACTATT TACGCAGAAT TICTAGGTGG GAGTTTCACT TGCGATGCCT ACCACATGAC
1030	TGGAATGAAG	1090	TCCCTTTTGT	1150	GGGATGGATG	1210	AATGAATGCT	1270	AGATGCGGTA	1330	GAGAAATTCC	1390	TATGGGGGAA	1450	TGCGACTATT

FIGURE 8 3/5

	٠																
1560	TGGCTCAGTC	1620	CTCCGGCTGG	1680	AGTTAAAAGT	1740	TGGAAGCAGT	1800	TGGAAAACCC	1860	TGAACGTTAA	1920	TCTTCGCCCC	1980	TTACATCTAG GACGTTTCGT GTGTGGAATT CTACTCAACA TATCAAAGCT GAAGTTTTGA	2040	CICCTIACGI CICIAGACAI GCCCAIGAGI ITIGIGICCG
1550	GAGAAGGCTT	1610	GCCACATCCA	1670	CAAAACAGAG	1730	ессеетеете	1790	GATCCATCCG AATATTAATT TGGAAAACCC	1850	GGGTCCTAAG AAGGAGAGAC TGAACGTTAA	1910	TCGTCCATAC	1970	TATCAAAGCT	2030	GCCCATGAGT
1540	CGAGCCTCAC CCTGATGGAG CTGGAGTGAT TCTCTGCATA GAGAAGGCTT TGGCTCAGTC	1600	AGGGAAGACG TAAATTACAT AAATGCCCAT GCCACATCCA CTCCGGCTGG	1660	CTCTTATCCA CTGTTTCGGC CAAAACAGAG AGTTAAAAGT	1720	AAATCAATGA TTGGTCACCT TCTCGGAGCA GCCGGTGGTG TGGAAGCAGT	1780		1840		1900	TGGGCACAAC	1960	CTACTCAACA	2020	CTCTAGACAT
1530	CTGGAGTGAT	1590	TAAATTACAT	1650	CTCTTATCCA	1710	TTGGTCACCT	1770	CAGGCAATAA GGACTGGGTG	1830	AATTGCTCGT	1.890	TTGGGTTTGG	1950	GTGTGGAATT	2010	
1520	CCTGATGGAG	1580	AGGGAAGACG	1640	AGATATCAAA GAGTACCAAG	1700	AAATCAATGA	1760		1820	AGATGAAGGC GTGGATACAA AATTGCTCGT	1880	GGTCGGTTTG TCTAATTCAT TTGGGTTTGG	1940	GACGTTTCGT	2000	ATGTTGGTAG
1510	CGAGCCTCAC	1570	AGGAGTCTCT	1630	AGATATCAAA	1690	TAATTCAACC	1750	TTCAGTAGTT	1810	AGATGAAGGC	1870	GGTCGGTTTG	1930	TTACATCTAG	1990	GGACTCCAGC

FIGURE

2100	ATACTCCTTG	2160	TGAAATCTCC	2220	CTCCTTGCAA TAGTTGTACT TTCGAGCTTT TCATCGAGTC AGTGAAGAAG AGAACAAAGC	2280	TCATCACCGT	2340	TTTTCTCAAA	
2090	GACACTTGAT	2150	TTTTTTCTC	2210	AGTGAAGAAG	2270	GCTCTCTATT	2330	GATTGGTTTG	·
2080	TACTCATGGC	2140	TCTCATATTT	2200	TCATCGAGTC	2260	CCTTTGTTTT	2320	GACTGGTTTA	ATCC
2070	ACGGATTGAG	2130	АТАТТСАТТА	2190	TTCGAGCTTT	2250	AACCATTTGC	2310	AAAACTAGAA	2370 GCTCTAGAGG
2060	GAGCTTTAGT CGGAACCATG ACGGATTGAG TACTCATGGC GACACTTGAT ATACTCCTTG	2120	CTAGAATTGT TGGTAGAGCA ATATTCATTA TCTCATATTT TTTTTTTCTC TGAAATCTCC	2180	TAGTTGTACT	2240	TGTTAACTCG GGCACGTAGT AACCATTTGC CCTTTGTTTT GCTCTCTATT TCATCACCGT	2300	TTTGTGGTTT TAAAATTTGT AAAACTAGAA GACTGGTTTA GATTGGTTTG	2360 AAGGGCGGCC
2050	GAGCTTTAGT	2110	CTAGAATTGT	2170	CTCCTTGCAA	2230	TGTTAACTCG	2290	TTTGTGTTT	2350 2360 2370 AAAAAAAAA AAGGGCGGCC GCTCTAGAGG ATCC

FIGURE 8 5/5

Sequence Range: 1 to 1580

666 G1y>	100	CAT TCG His Ser>	150	GTG Val>		GAT ASD>		TCT Ser>	AAA Lys>	340	CGC Arg>
50 TCT Ser	ਜ			AGG Arg		GGT		GGA	290 GCT Ala	ň	ATC Ile
GCA Ala		CAG Gln		AAA Lys	190	TTG	240	ATT Ile	CTT CEU		GGG
AAT Asn		ACT Thr	140	TCC	13	TCT		TTA Leu	GAT Asp		ACG
40 GCG Ala	90	GCA	П	GTC Val		CAG Gln		AAA Lys	280 AAT GAT Asn Asp	330	CGA Arg
40 ATG GCG Met Ala		AGG Arg		$ ext{rTT}$		agg Arg	230	TGC	28 AAT Asn		GTC Val
ളള്ള		CTG AGA Leu Arg	130	GAG Glu	180	GAC	(4	GGA Gly	TCA		ACT
g d	80	CTG	13	TCG		TCT		AGA Arg	GTC Val	320	ATT
3(CGT		GCC		TCC		gat Asp	220	AGT Ser	270 CAA Gln	(*)	TGG
4GTT)		CCT		TCT	170	CAG Gln	2	GTG Val	CTT Leu		GAA
02 84 85 84 87 84 84 84 84 84 84 84 84 84 84 84 84 84	70	GTT Val	120	GGA G1y	7	GTT Val		CTT	GCT	0	GAT
AAGAC		TCA		CGT Arg		GCC		AGG Arg	260 CCA Pro	310	AAT
10 CCTGAATCGG ATTCAAGAGA GAGTTTCGTT GCTGGG		TCT		TCT	160	AGT	210	CCG	2 ATA Ile		ACC
10 366 7		GGT	110	TCG	16	TGT Cys		TCG	GCT		GAC
SAAT	6 0	CTG	••	TCA		TGC Cys		CGC	o TCT Ser	300	GTC
CCTC		TTT Phe		ATT Ile		TTT Phe	200	TCT	250 GGT TCT (Gly Ser A		ATT

FIGURE 9 1/5

					,			_				
390	TCA Ser>		GAT Asp>		GGC Gly>	TTG Leu>	580	GTC Val>	630	GTG Val>		GGA G1y>
	GCA		AAT Asn		TTC	330 CCT P	$\tilde{\Omega}$	TTA		CTA		CGG Arg
	TTA Leu	430	GCA Ala	480	CTT	S AAT Asn		GGT		ATT Ile	029	GAT Asp
380	AAT TTA Asn Leu	43	GAC		GAC	AAG Lys		TTG	620	AAT Asn	67	ACC
(7)	ACA Thr		GTA		GAG Glu	20 AAA Lys	570	GTG Val	w	AAC Asn		TGG Trp
	CTT Leu		CAG Gln	470	CCT	520 TGC AAA		TTT Phe		TTT Phe		GAC
370	AGT	420	GCA Ala	4	ACC	66C 61Y		GGA Gly	0,	GGT	660	GTT Val
37	GAT		ATG Met		TCT	CTT Leu	260	AGT	610	GGG G1y		TAT
	AAA Lys		GAG Glu	460	ACT	510 GCA Ala	u)	TGC		GGT		CGG
	TCA GGT Ser Gly	410	CTA	46	TGT	AAA Lys		GCA Ala		AGA Arg	029	TCT
360	TCA	7	GCT Ala		ATG	500 ATA TCG Ile Ser	550	GCT	009	ATT Ile	•	CTT
	CTC		AAA Lys		TTG	500 ATA Ile	55	ACC		CAC		TCT
	GTT Val	400	AGG	450	GTT Val	cag Gln		ATT		TGC	640	GAT
350	agg Arg	4(GCA		ATG	CCT		GAC	590	GCT	9	GCT Ala
•••	CGA Arg		GCA	7	GAT	490 T GCT r Ala	540	TAC	u ;	GCT		GGT
	AAC		GAG Glu	440	GTG Val	49 AGT Ser		TCT		TCA		ATT Ile

FIGURE 9 2/5

	TCA Ser>	GAT Asp>	820	GTT Val>	870	AGG Arg>		CGC Arg>		AAG Lys>	GCA Ala>
	CAG Gln	770 AGC	88	GAA		CCA		TTC Phe		GGA Gly	1010 T CAT CAG tu His Gln
720	GTG Val	CAT His		GAT Asp		CCA	910	GTA Val	960	CTT	1(CAT His
	GTG	TTG		GAA Glu	860	TTT Phe	9	GAG Glu		요설	្ន្រី
٠	GTA Val	50 GAT ASP	810	AAA Lys	w	GAT Asp		AAA Lys		TCA	CTG CTG Leu
710	GCT	760 TTT GA Phe As		ATC Ile		aga Arg		GGT Gly	950	GAA Glu	1000 TTG CT Leu Le
	GGA Gly	GCT		GCA Ala	850	ATC	900	AAC Asn	O1	ATC Ile	TGG
	GCT	TTT Phe	800	GCT Ala	8	TCC		ATG Met		TCA	GAC Asp
700	GCT	750 CTC Leu	w	AAA Lys		$^{\rm GGG}_{\rm G1Y}$		CAA Gln	940	CAG Gln	990 ATC Ile
7(GAT Asp	666 G1y		CTA		AAT	890	ATC Ile	6	CCT	AAC Asn
	GGA Gly	GAT Asp	190	CAT His	840	CAT	w	TGC		GTG Val	TCC
	TTT Phe	740 GAA Glu	7.5	AGG		GGA G1y		TCT		TCT	980 GGA G1y
069	CTC	7 GAG Glu		CAA		CTG	880	TAC	930	CGC	9 AAT Asn
	ATT	GCT		GGG G1y	830	GCC	88	TCA		TGC	CTT Leu
	TGT Cys	730 T GAT	780	GAT ASP	ω	AAA Lys		TCT		GCT	70 GGT Gly
089	ACA	73 TGT Cys		GGA Gly		GAT Asp		CGT	920	TTT Phe	970 GCC GC Ala G

FIGURE 9 3/5

09	CCT CAA Pro Gln>	1110	GCA Ala>		AAG Lys>		TGG Trp>	1260	CACTGCAGCT	1320	TCCTCTCAAA CCGATGTTTC ACGAAATTTT GCTTCCATGA CCANAAAAAG AAGAAGTCAG	1380	AGCAAGCAAC ACGACACGAT CTTCATCACA TTGCCCTTTT TCGTTCCCCT
1060	CCT		GCG		GTG		ACA		CACT		AAGA		TCGT'
	GTT Val		AGT	1150	AAT	1200	CTC	1250		1310	AAG	1370	TTT
	gag Glu	1100	ACT	11	GGA		GGA G1y	7	GAGC	Н	NAAA	-	CCCT
1050	CTA	Н	AAC		AGT		GCC) (3)		CCA		TTG(
	CGT		AAT TAC GGG Asn Tyr Gly		AGG Arg	1190	TTT GGC Phe Gly	1240	GGA TAA GACTGAA GCCGAGCCAG Gly ***>	1300	ATGA	1360	CACA
	GCA ACA Ala Thr	0	AAT TAC Asn Tyr	1140	GTG Val	ਜ	$ extsf{TT}$	••	GAC	••	TCC	• •	CAT
1040	GCA	1090	AAT Asn	• •	GCT		GGA Gly		TAA (_	ည်	_	CF
ਜ	GCA GTA (GCA		CTA GAC GAA GCT GTG AGG Leu Asp Glu Ala Val Arg	0	GCA Ala	1230		1290	TTT	1350	CGAT
	GCA		TTG Leu	1130	GAC	1180	ACC	-	TGG Trp		GAAA		GACA
0	gat Asp	1080	TCA AAC Ser Asn	11	CTA		GCA Ala		AGG Arg	0	C AC	0	C AC
1030	ATT	П	TCA		GCA		ATT Ile	1220	ATC	1280	GTTT	1340	GCAA
	ATC Ile		ATC Ile	0	CCC TTG Pro Leu	1170	GTG Val	12	ATT ATC Ile Ile		CGAT		GCAA
	AGG	1070	ATT Ile	1120	CCC	-	CAC His		GCT	1270	AA C	1330	
1020	cag agg Gln Arg	10	CGA Arg		ATT Ile		GGT	0.	TCT	12	CTCA	13	TTAT
н	AAT Asn		GAA Glu		TCC	1160	CCG	1210	GGT G1y		TCCT		TCTTTTATGG

FIGURE 9

1500 * CGGGACATTG 1560 * AAAAAAAAA	1490 CATTTTGTCT 1550 AAAAAAAAA	1450 1460 1470 1480 1490 1500 * TAAGTTATTT GTTTCTTGTT TAATTGTTCA GCTTTTTACTT CATTTTGTCT CGGGACATTG 1510 1520 1530 1540 1550 1560 * GAGATGACAG CATAAACATC ATGTTTATAT TTTGCTAAAA AAAAAAAAA 1570 1580 AAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA	1470 TAATTGTTCA 1530 ATGTTTATAT	1450 1460 TAAGTTATTT GTTTCTTGTT 1510 1520 GAGATGACAG CATAAACATC 1570 1580 AAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA	1450 ATTT G 1510 ACAG C ACAG C
15 AAAAAAA	1550	1540 TTTGCTAAAA	1530 ATGTTTATAT	1520 CATC	CATAAA
150 CGGGACAT1	1490 CATTTTGTCT	1480 GCTTTTACTT	1470 TAATTGTTCA	1460 TGTT	GTTTCI
TTGTCCCCA	ATAGTTTCTT	TITCCATTAG TITGATGATT TIGCTGACAA TACAATACCC ATAGTTTCTT TIGTCCCCAA	TTGCTGACAA	GATT	TTTGAT
1440	1430	1420	1410	1400	14

FIGURE 9 5/5

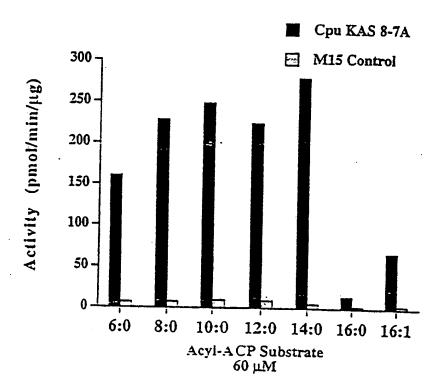


FIGURE 10

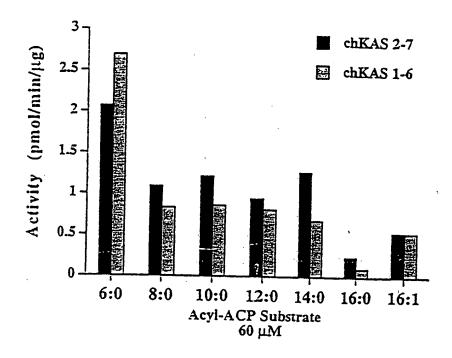


FIGURE 11

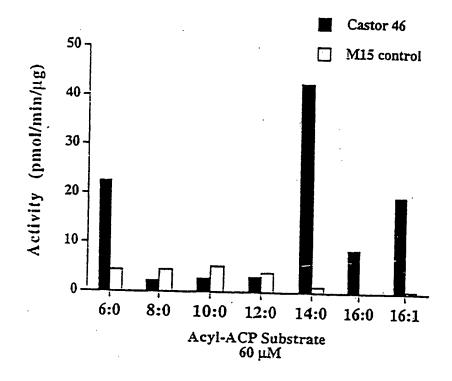
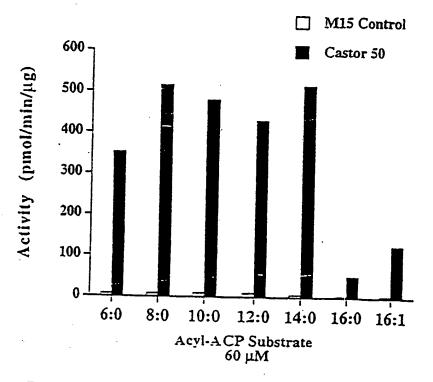


FIGURE 12



E328013-28

FIGURE 13

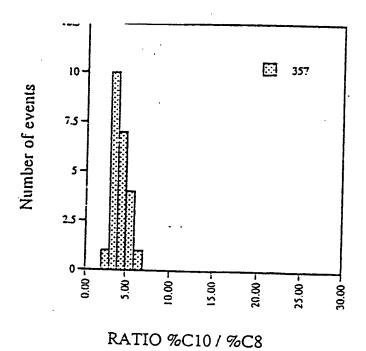


FIGURE 15

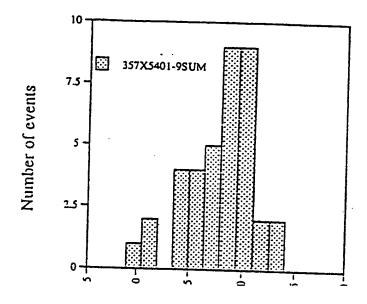
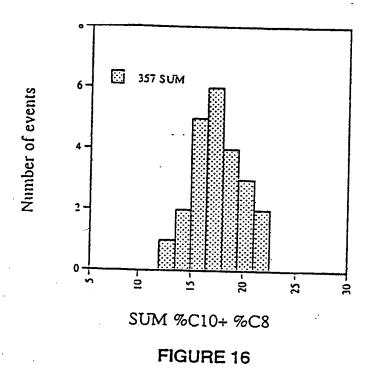


FIGURE 15 2/2



SUBSTITUTE SHEET (RULE 26)

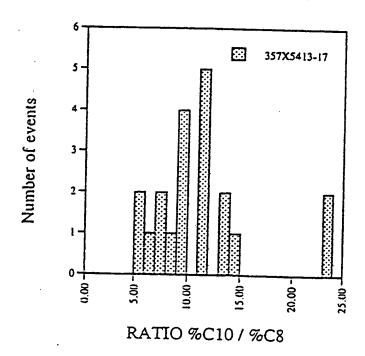


FIGURE 17 1/2

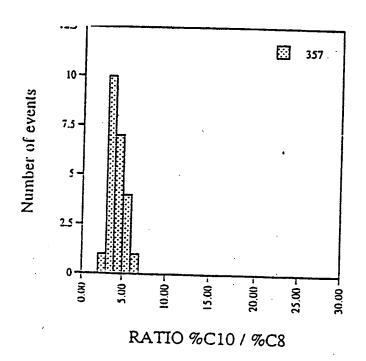


FIGURE 17

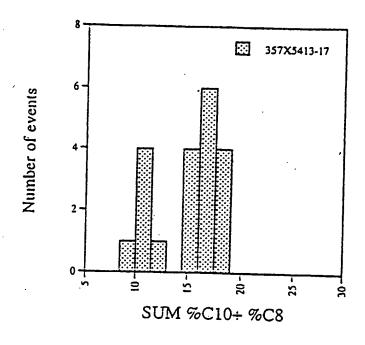
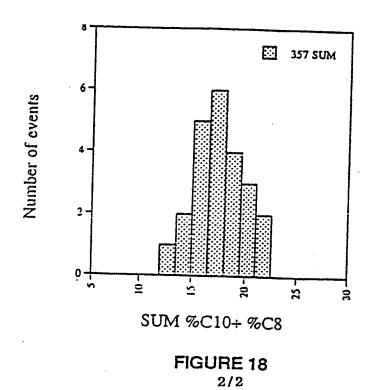
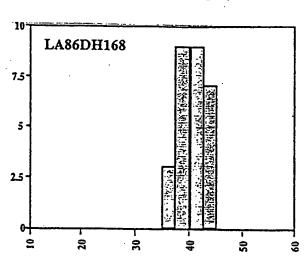


FIGURE 18 1/2



SUBSTITUTE SHEET (RULE 26)





12:0 levels (w%)

FIGURE 19 1/3



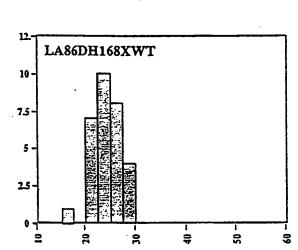
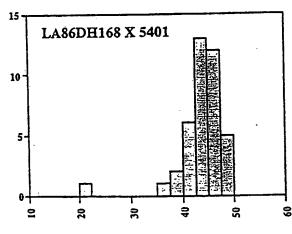


FIGURE 19 3/3 SUBSTITUTE SHEET (RULE 26)



12:0 levels (w%)

FIGURE 19 2/3.

SUBSTITUTE SHEET (RULE 26)

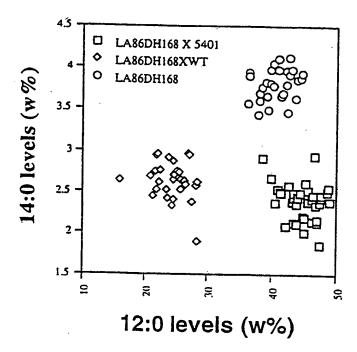
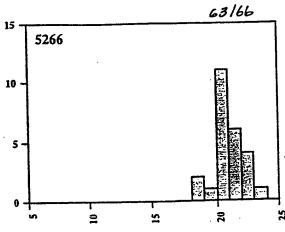


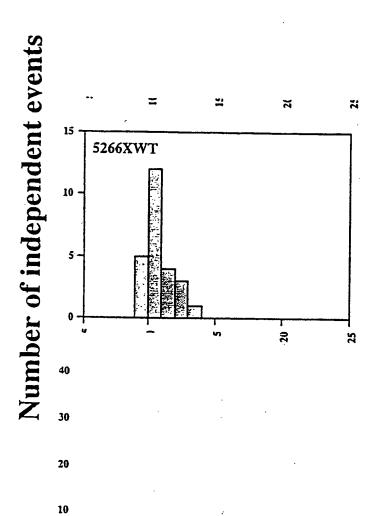
FIGURE 20





18:0 levels (w%)

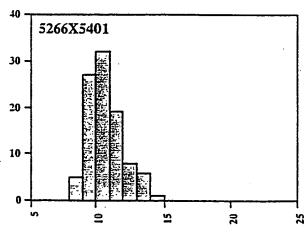
FIGURE -21. 1/3



18:0 levels (w%)

FIGURE 21. 2/3

Number of independent events



18:0 levels (w%)

FIGURE 21

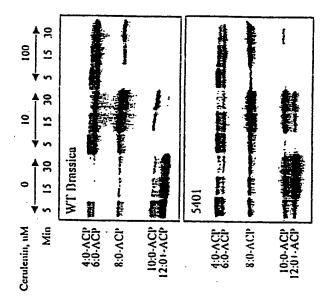


FIGURE 22

PCT

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(74) Agent: SCHWEDLER, Carl, J.; Calgene LLC, 1 Street, Davis, CA 95616 (US).	920 Fi	fth .
·		
(54) Title: PLANT PATTY ACID SYNTHASES AND	USE IN	IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN

(54) Title: PLANT PATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

(57) Abstract

By this invention, compositions and methods of use related to β -ketoacyl-ACP synthase of special interest are synthases obtainable from *Cuphea* species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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CN	China	KR	Republic of Korea	PT	Portugal		
CU	Cuba	KZ	Kazakstan	RO	Romania		
cz	Czech Republic	LC	Saint Lucia	RU	Russian Federation		
DE	Germany	LI	Liechtenstein	SD	Sudan		
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Inte. Jonal Application No PCT/US 98/07114

			01/03 90/0/114
A. CLASSIF IPC 6	FICATION OF SUBJECT MATTER C12N15/82 C12N15/54		·
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Documentat	ion searched other than minimum documentation to the extent that s	uch documents are included	i in the fields searched
Electronic da	ata base consulted during the international search (name of data ba	se and, where practical, se	arch terms used)
C. DOCUME	ENTS CONSIDERED TO BE RELEVANT		
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х	WO 95 06740 A (MAX PLANCK GESELL; TOEPFER REINHARD (DE); MARTINI (D) 9 March 1995 see page 16, paragraph 1; claim	NORBERT	15,22
P,X	LEONARD, J.M., ET AL.: "A Cuphe beta-ketoacyl-ACP synthase shift synthesis of fatty acids towards chains in Arabidopsis seeds expr Cuphea FatB thioesterases" THE PLANT JOURNAL, vol. 13, no. 5, March 1998, page XP002081429 see the whole document ———	s the shorter essing	15,22, 29-32
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	actual completion of theinternational search	Date of mailing of the	international search report
2	0 October 1998	02/11/19	98
Name and	mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2	Authorized officer	
	NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Maddox,	A

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C.(Continu	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	
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Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
·
2. X Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically: See FURTHER INFORMATION sheet
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of Invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Claims Nos.: 1-14,19,20,21,26,27,28

Remark: Claims 1-14 were not provided to the ISA at the time of search and hence the subject matter of these claims and the dependent claims 19,20, 21,26,27,28, could not be defined.

Information on patent family members

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